

Coordination Polymer Nanoparticles: New Objects and New Trends.

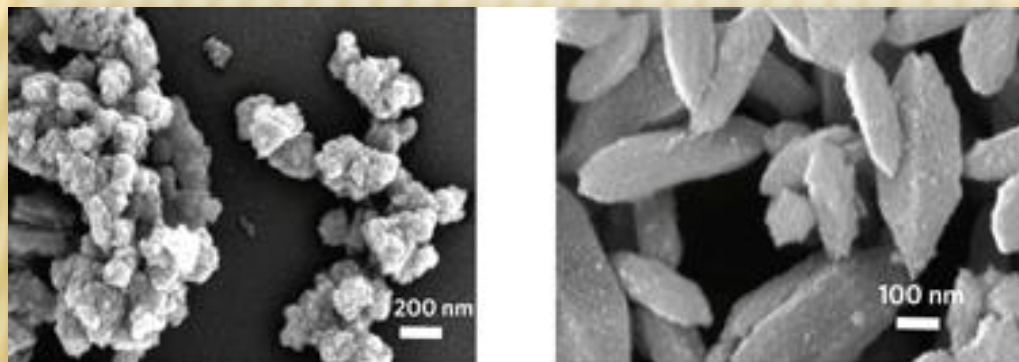
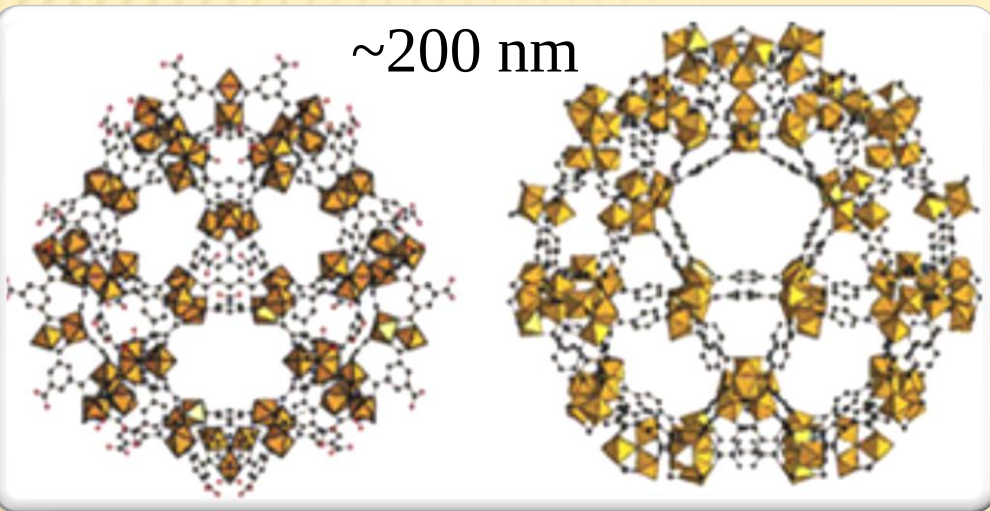
Joulia Larionova

- A new type of inorganic nanoparticles
- Multifunctional magneto-optical nanoparticles
- New contrast agents for MRI and biolabeling

A New type of Inorganic Nanoparticles: *Coordination Polymer Nanoparticles*

Porous Iron Carboxylates Frameworks Nanoparticles

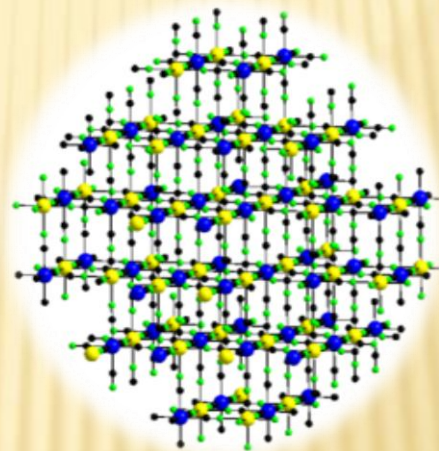
~200 nm



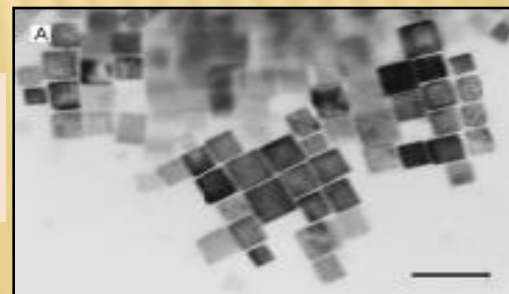
G. Ferey, *Nature Materials* 2010, 9, 172

Cyano-Bridged Metallic Nanoparticles

1 - 100 nm



Nanocubs of
 $\text{Fe}^{\text{III}}_4[\text{Fe}^{\text{II}}(\text{CN})_6]_3$



S. Mann, *Angew. Chem. Int. Ed.*, 2000, 39, 1793

Cyano-Bridged Metallic Coordination Polymer Nanoparticles

The oldest bulk coordination polymers...

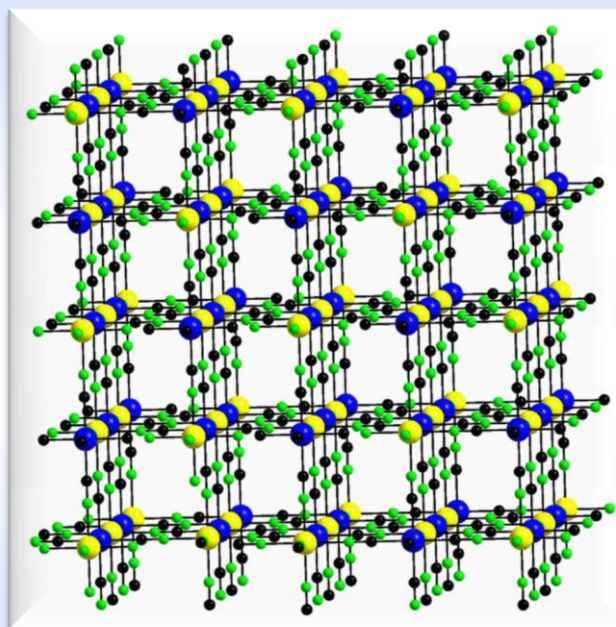
Prussian Blue:



Discovered in 1704



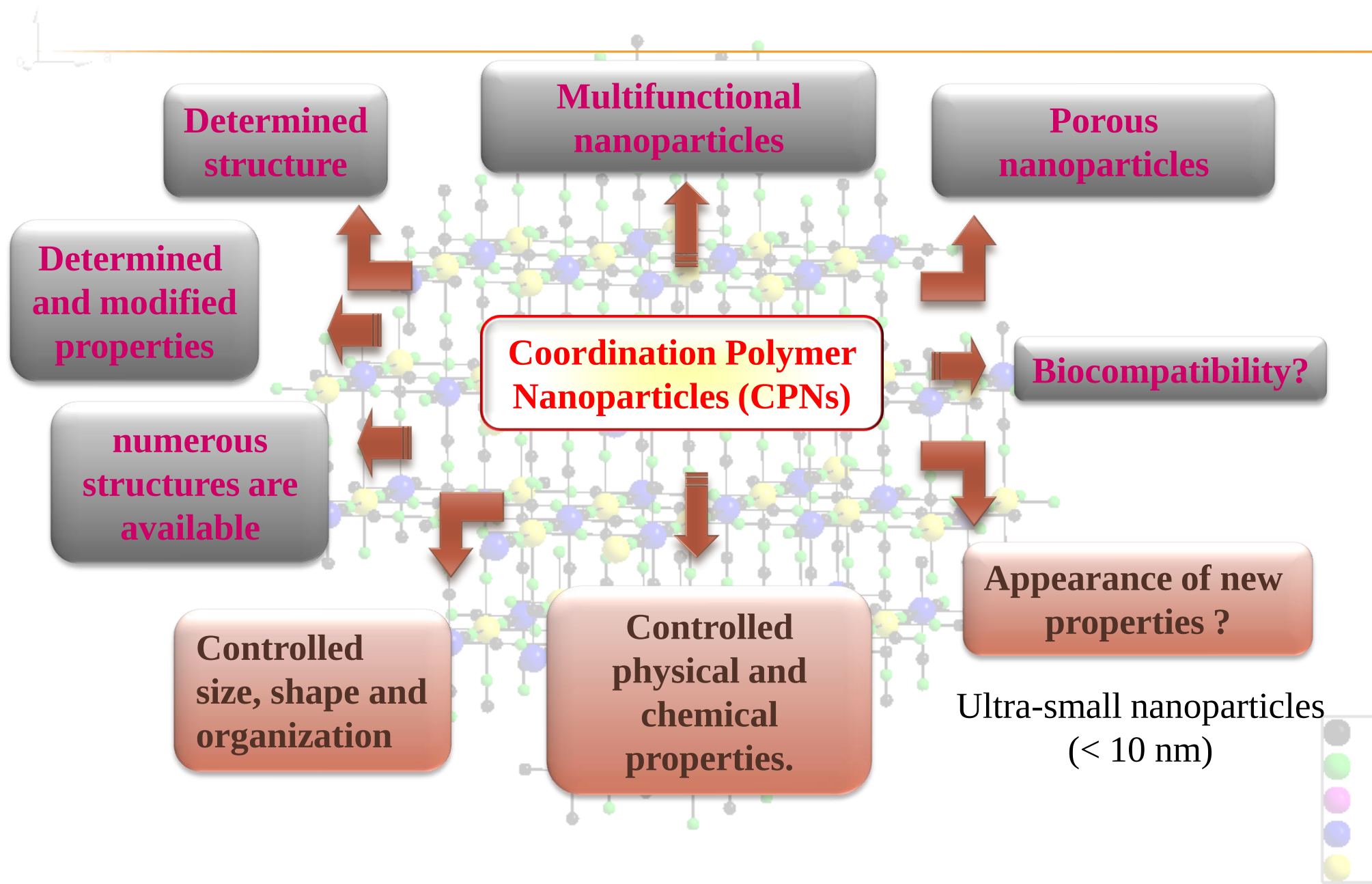
Johann Conrad Dippel



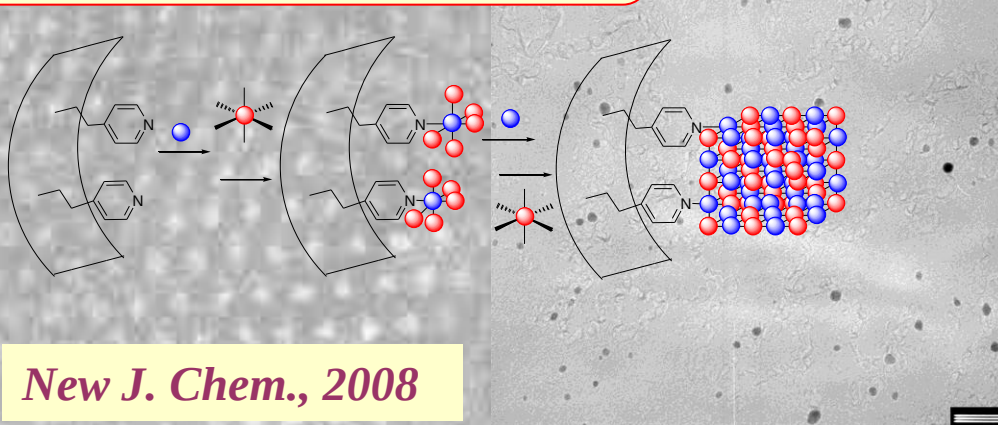
M and M' = transition
metal ions or lanthanides

- a pigment;
- magnetic properties: molecular magnets;
- photo-magnetic properties;
- host-guest properties: molecular sieves;

Cyano-Bridged Metallic Coordination Polymer Nanoparticles

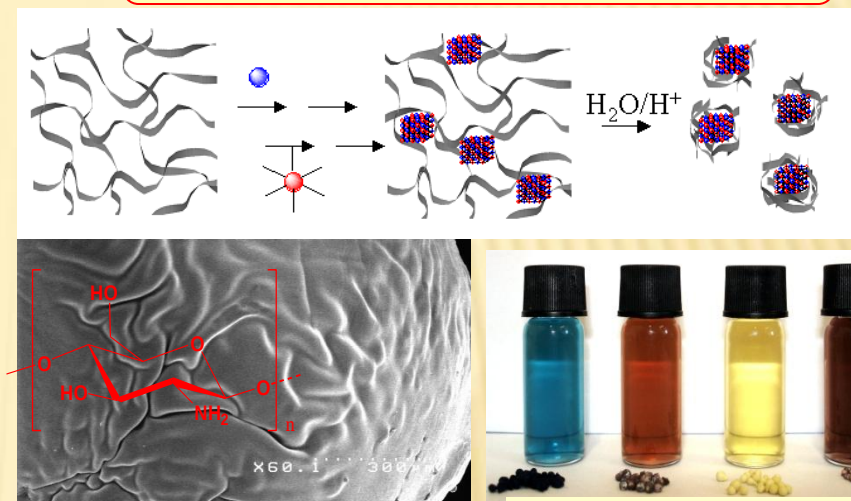


Nanoparticles into mesostructured silica



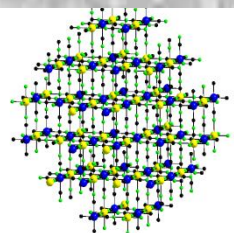
New J. Chem., 2008

Nanoparticles into chitosan

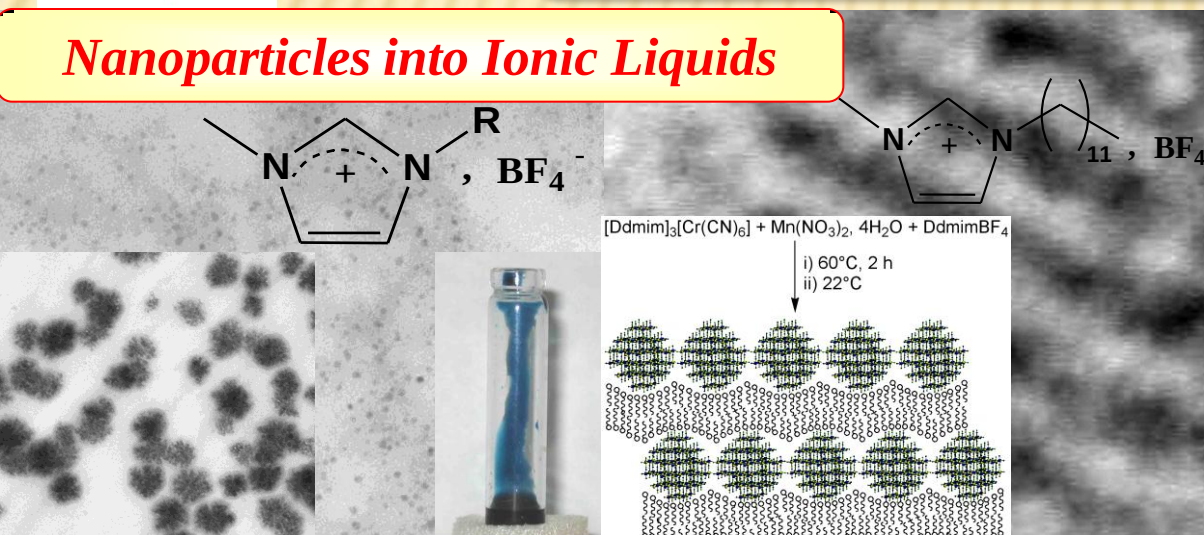


Chem Commun. 2006;
Dalton Trans. 2008
Angew. Chem. 2009

Nanoparticles of cyano-bridged coordination polymers

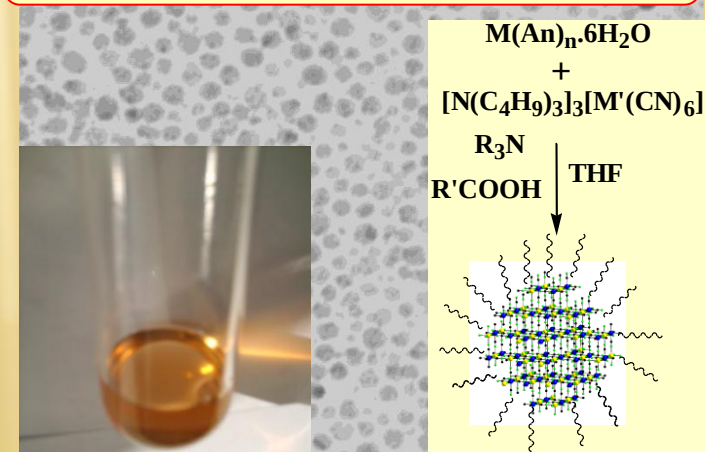


Nanoparticles into Ionic Liquids



Chem: Eur. J., 2006; Inorg. Chim. Acta 2007, 2008

Nanoparticles with Ligands



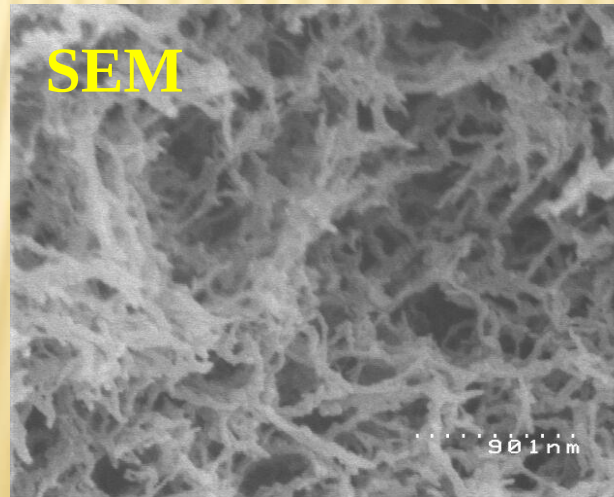
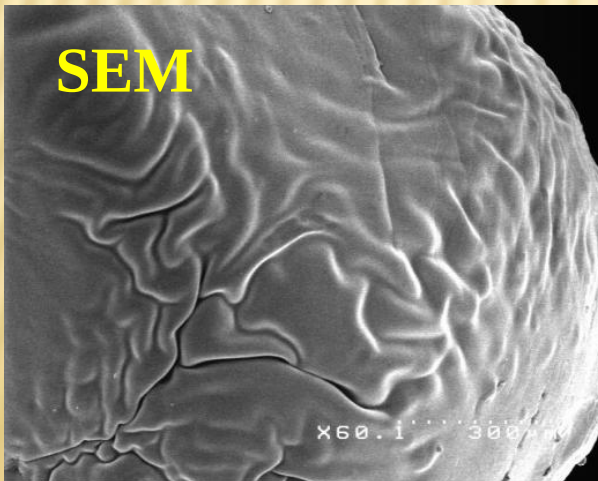
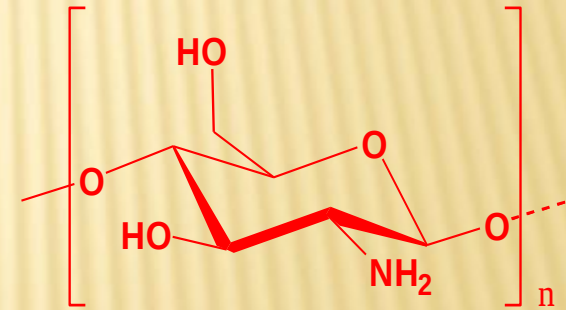
Chem.Mater., 2007

Water Soluble Cyano-Bridged Metallic Nanoparticles enwrapped by biopolymer chitosan

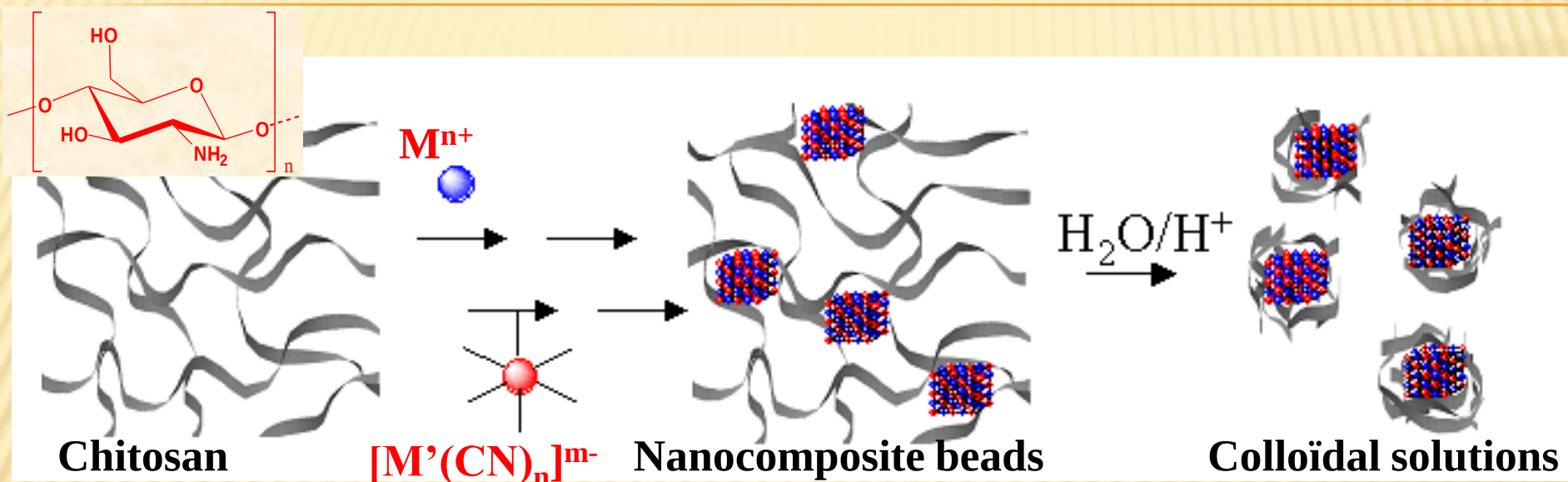


Why we use the chitosan beads?

- a porous structure - diffusion of precursors;
- NH_2 groups able to coordinate metal ions -
- high water solubility;
- biocompatibility.



Water Soluble Cyano-Bridged Metallic Nanoparticles enwrapped by biopolymer chitosan



$M^{2+} = \text{Fe, Ni, Co, Mn, Cu...}$ and

$M^{3+} = \text{Gd, Yb, Eu...}$

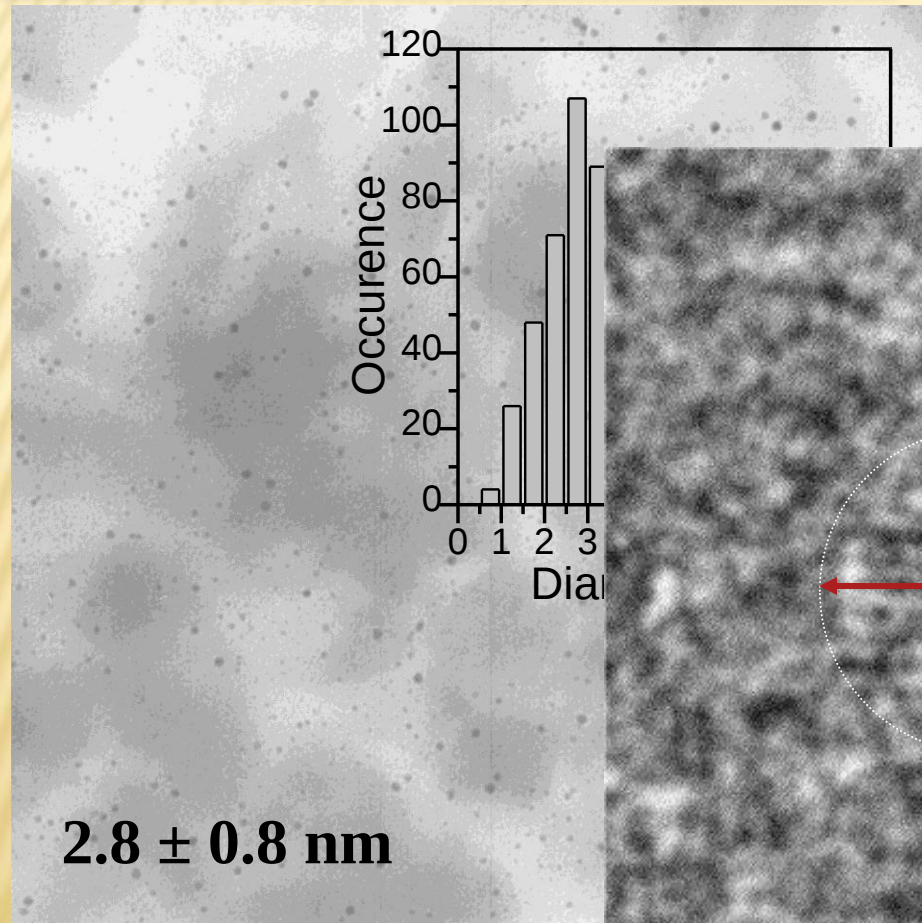
$M' = \text{Fe, Cr, Co (n = 6); Ni, Pt (n = 4), Mo (n = 8)}$

Chem Commun. 2006;
Dalton Trans. 2008,
Angew. Chem. 2009,
PCCP 2010

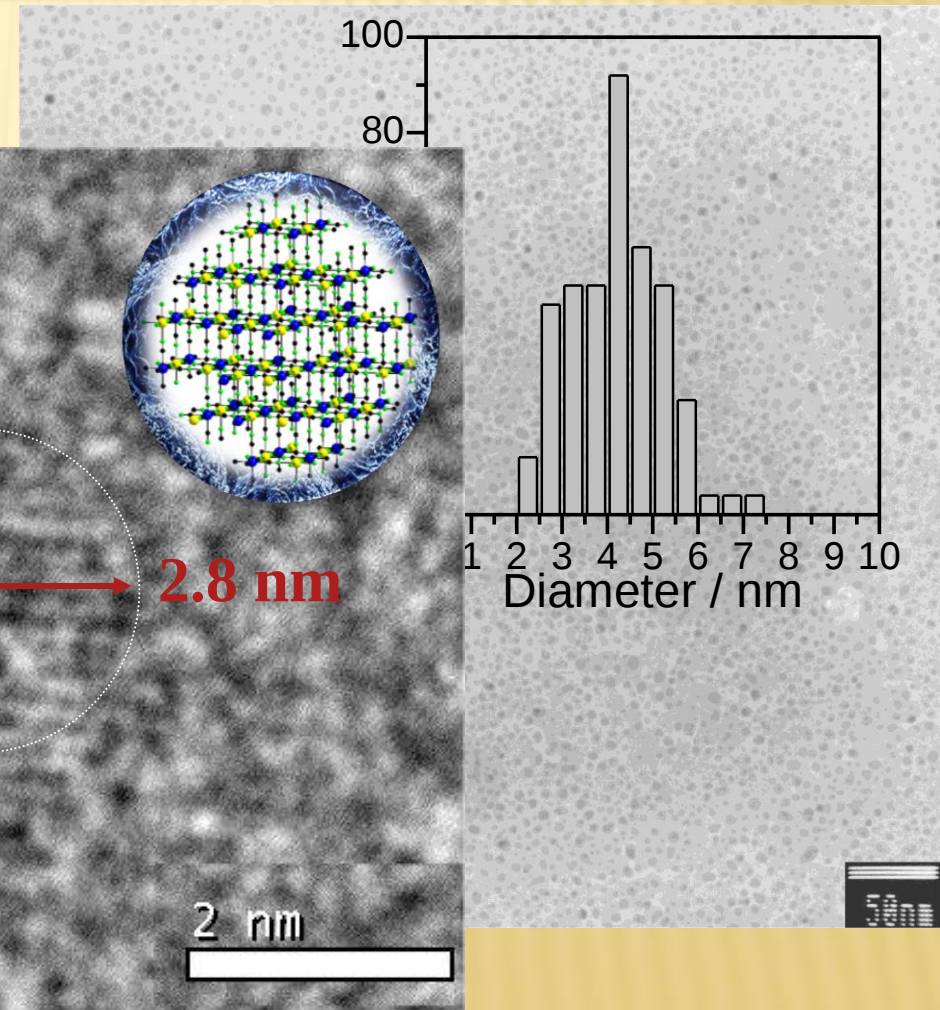


$\text{Ni}^{2+}/[\text{Fe}(\text{CN})_6]^{3-}/\text{chitosan}$ Nanoparticles

Beads of $\text{Ni}^{2+}/[\text{Fe}(\text{CN})_6]^{3-}/\text{chitosan}$



Aqueous colloid of $\text{Ni}^{2+}/[\text{Fe}(\text{CN})_6]^{3-}/\text{chitosan}$

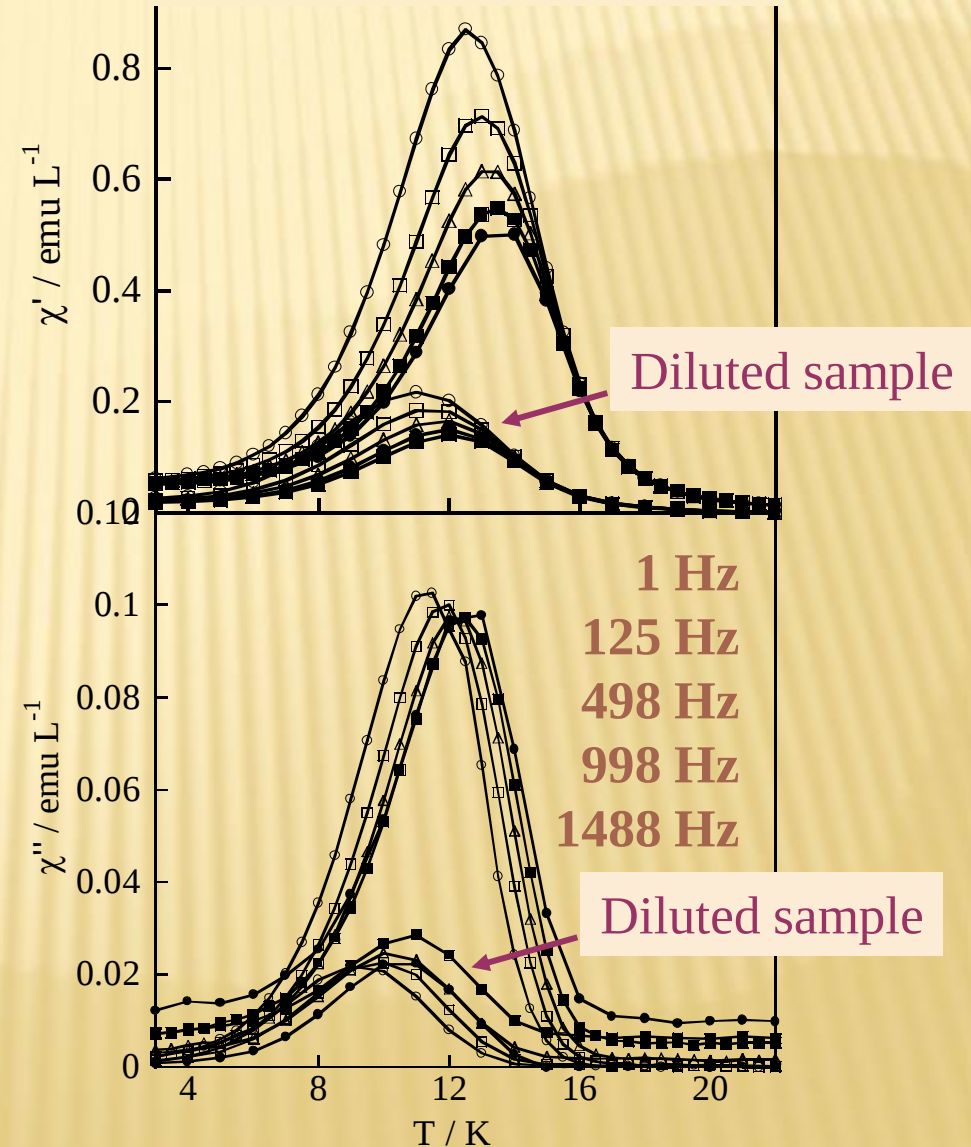
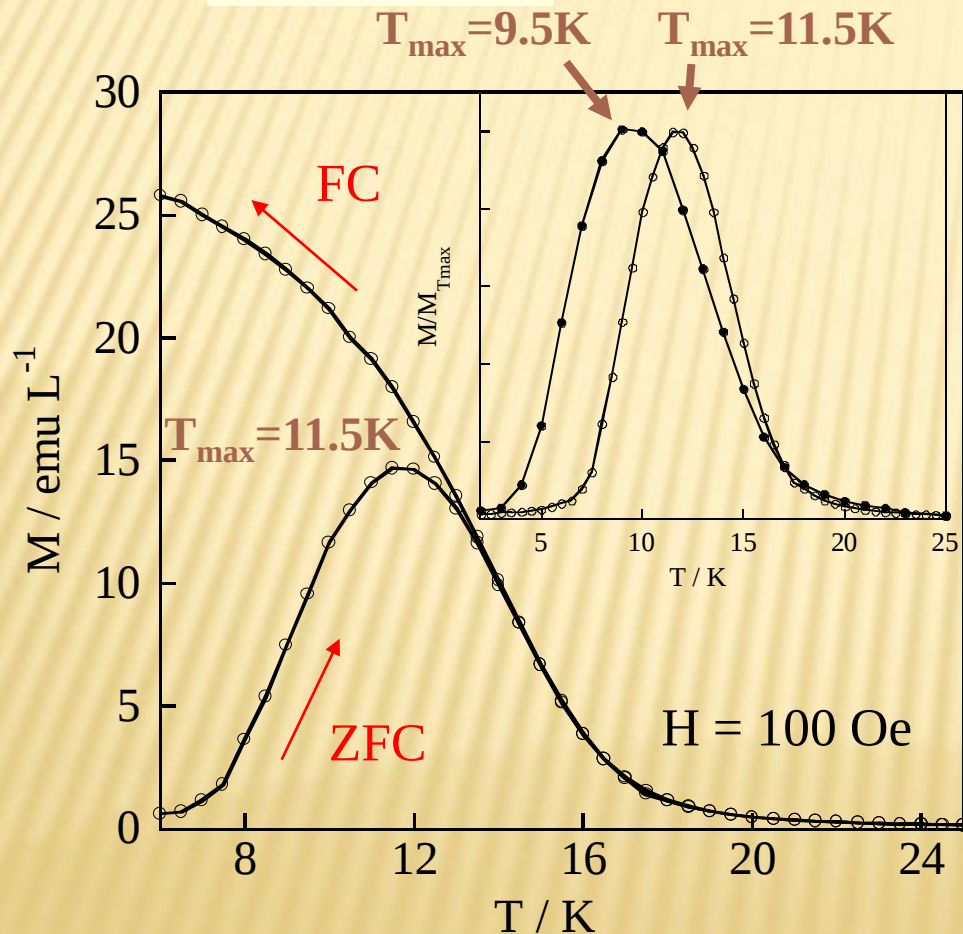


Magnetic Properties as a probe to study nanoparticles behavior

1 *Dynamic analysis:*

Colloids $\text{Ni}^{2+}/[\text{Fe}(\text{CN})_6]^{3-}/\text{chitosan}$

Diluted sample:



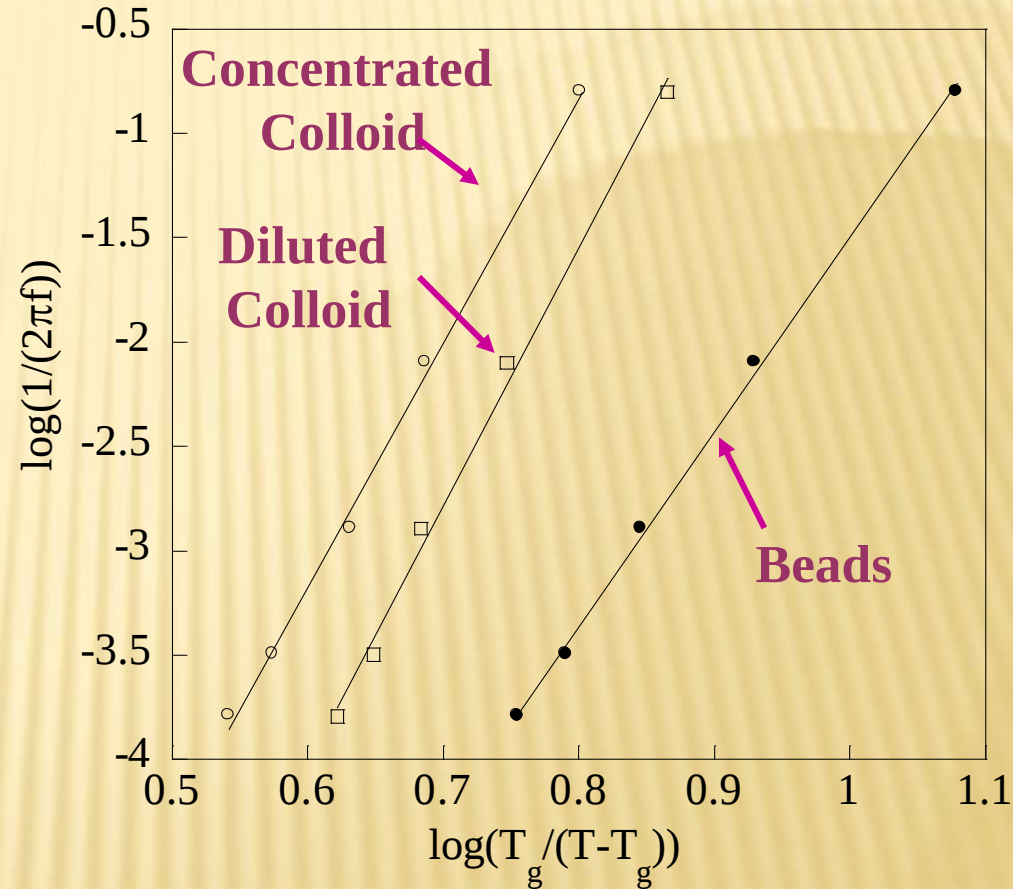
Magnetic Properties as a probe to study nanoparticles behavior

Critical slowing down:

$$\tau = \tau_0 [T_g / (T_{max} - T_g)]^{z\nu}$$

T_g , K	τ_0 , s	$z\nu$
Colloids $\text{Ni}^{2+}/[\text{Fe}(\text{CN})_6]^{3-}/\text{chitosan}$		
11.2	$3.56 \cdot 10^{-12}$	12.4
Diluted $\text{Ni}^{2+}/[\text{Fe}(\text{CN})_6]^{3-}/\text{chitosan}$		
9.5	$7.29 \cdot 10^{-11}$	11.7

$$z\nu = 4 - 12; \tau_0 = 10^{-8} - 10^{-13} \text{ s}$$

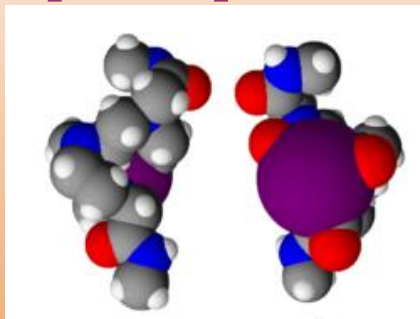
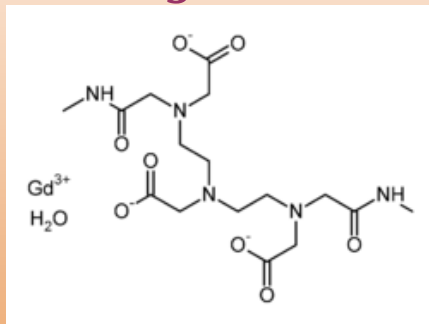


Superspin-glass behavior due to the presence of interparticle interactions and surface spin frustration

Coordination Polymer Nanoparticles as Contrast Agent for Magnetic Resonance Imaging:

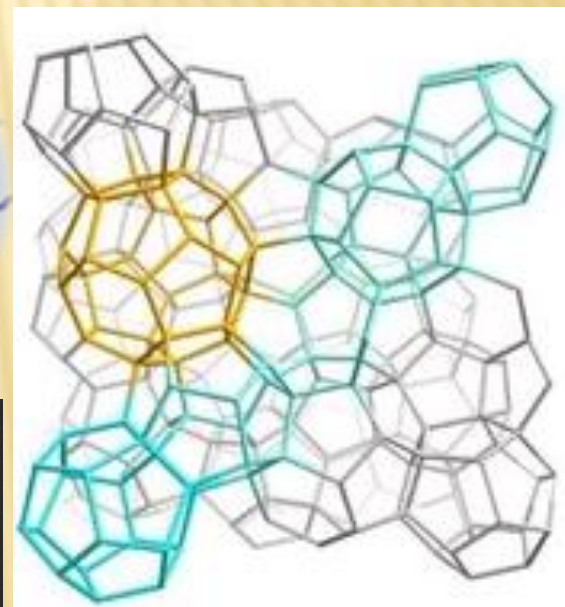


Paramagnetic Gd^{3+} complexes: positives CA

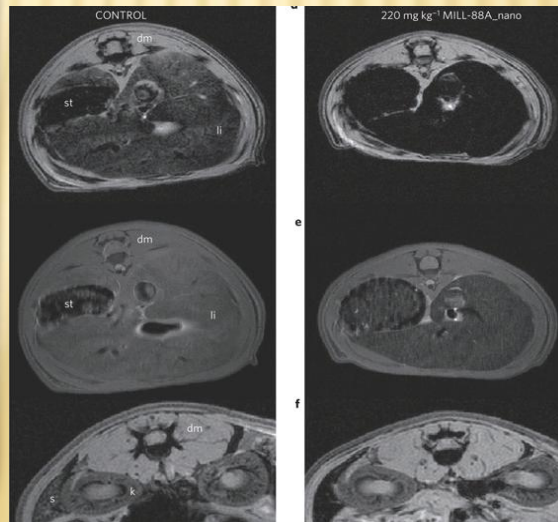
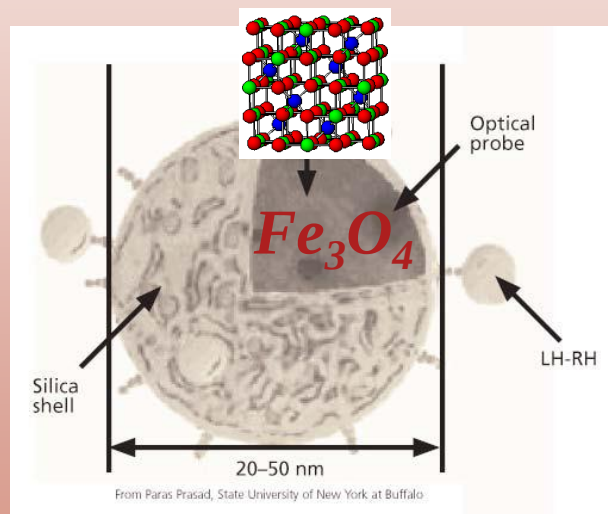


Omniscan®, Gadoteridol®, Dotarem®...

Porous Iron Carboxylates Frameworks Nanoparticles



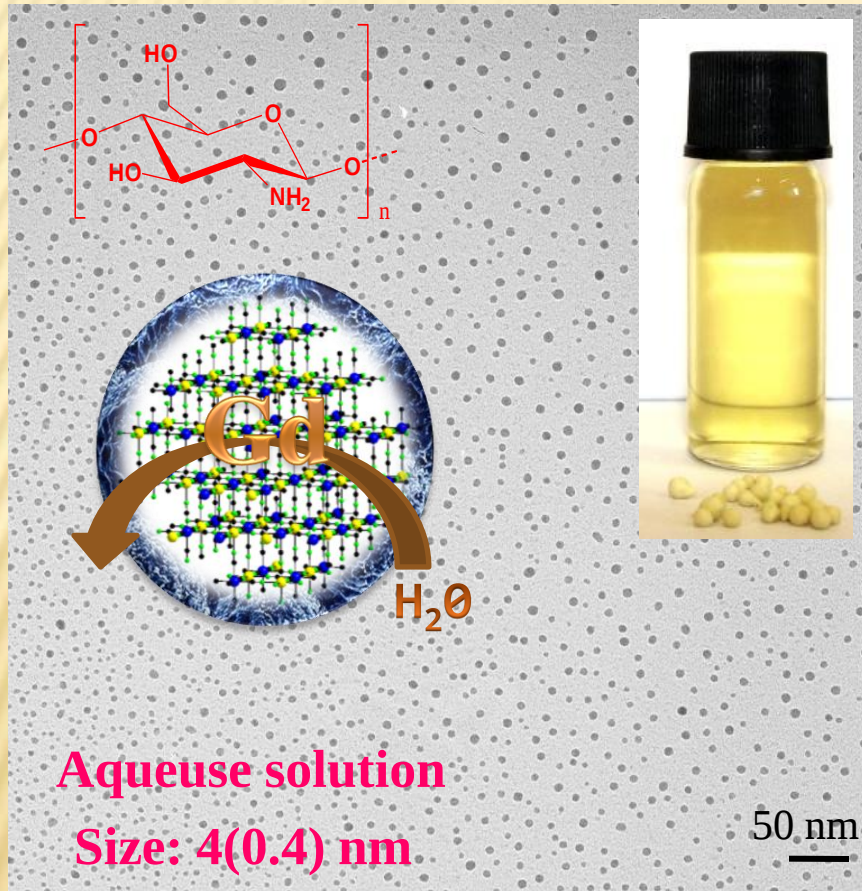
SPIONs: negatives CA



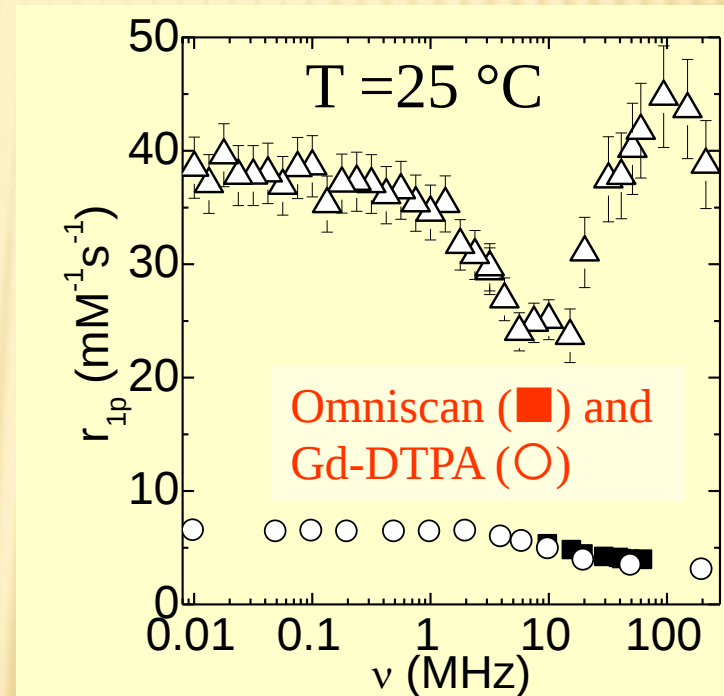
Resovist®, Combidex®, Endorem®...

G. Ferey, *Nature Materials*
2010, 9, 172

Gd[Fe(CN)₆]/chitosan Nanoparticles



Longitudinal relaxivity



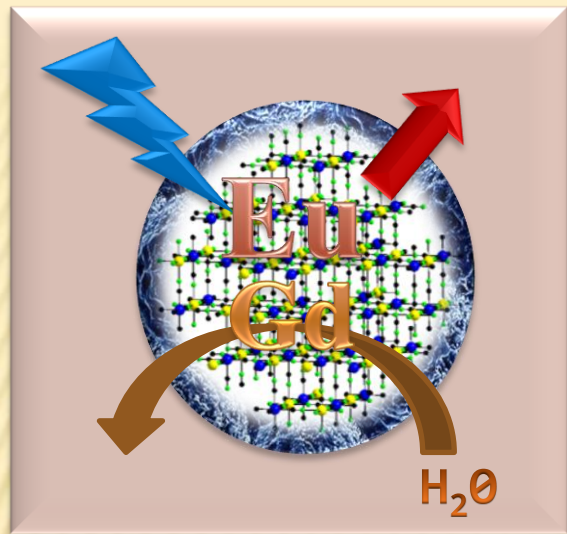
$$r_{ip} = [(1/T_i)_{\text{meas}} - (1/T_i)_{\text{dia}}] / c \quad i=1,2$$

$(1/T_i)_{\text{meas}}$ is the measured value on the sample with concentration $c = 4 \text{ mmol L}^{-1}$ of Gd^{3+}
 $(1/T_i)_{\text{dia}}$ is the nuclear relaxation rate of water

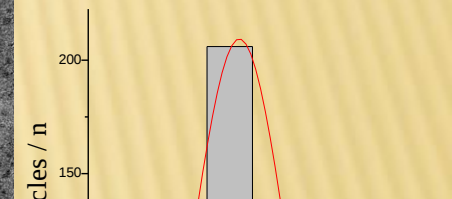
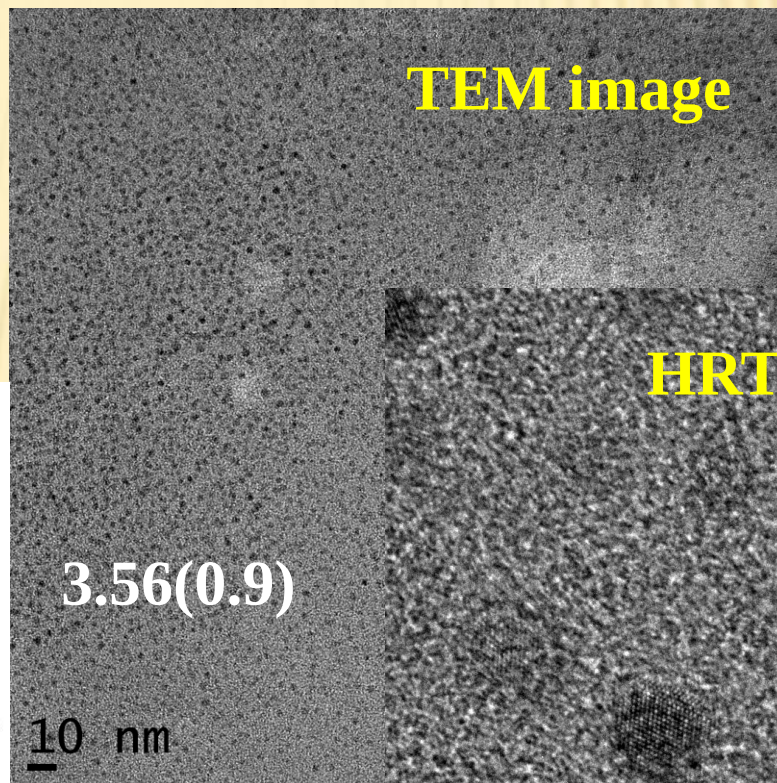
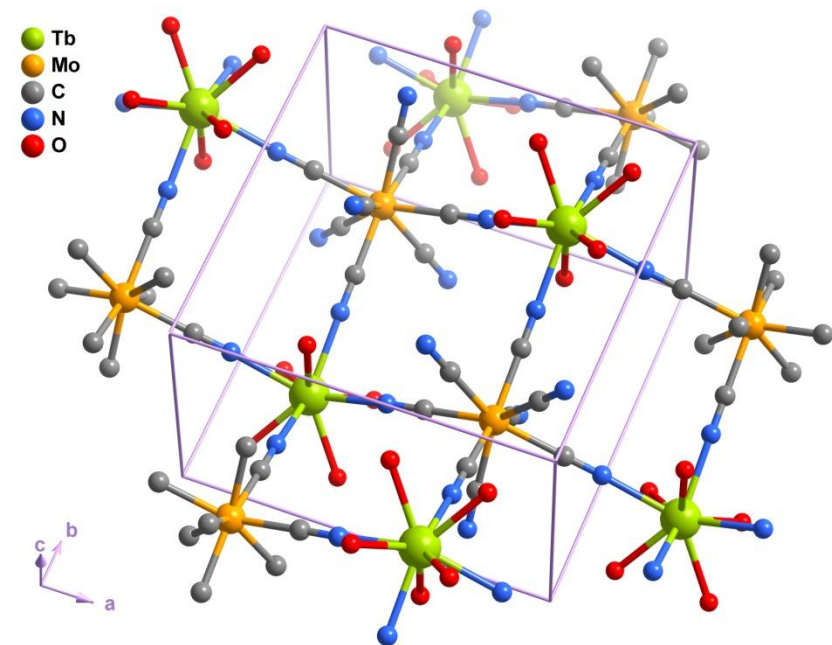
Dalton Trans. 2008.

Multifunctional $\text{Gd}_{0.5}\text{Ln}_{0.5}[\text{Mo}(\text{CN})_8]/\text{chitosan}$ (Ln = Eu, Tb) Nanoparticles

Aqueous colloidal solutions:

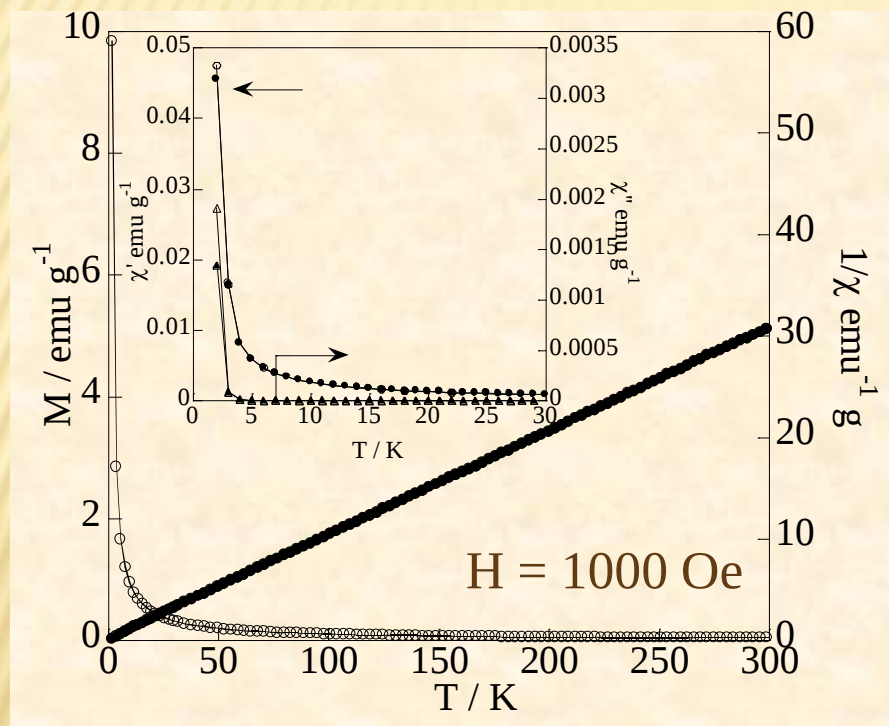


Crystal structure of bulk
 $\text{Gd}_{0.5}\text{Ln}_{0.5}[\text{Mo}(\text{CN})_8]$

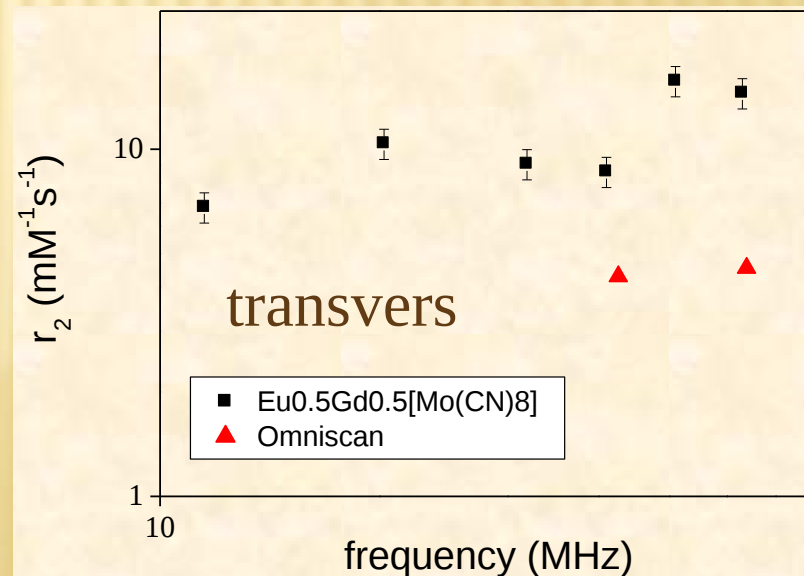
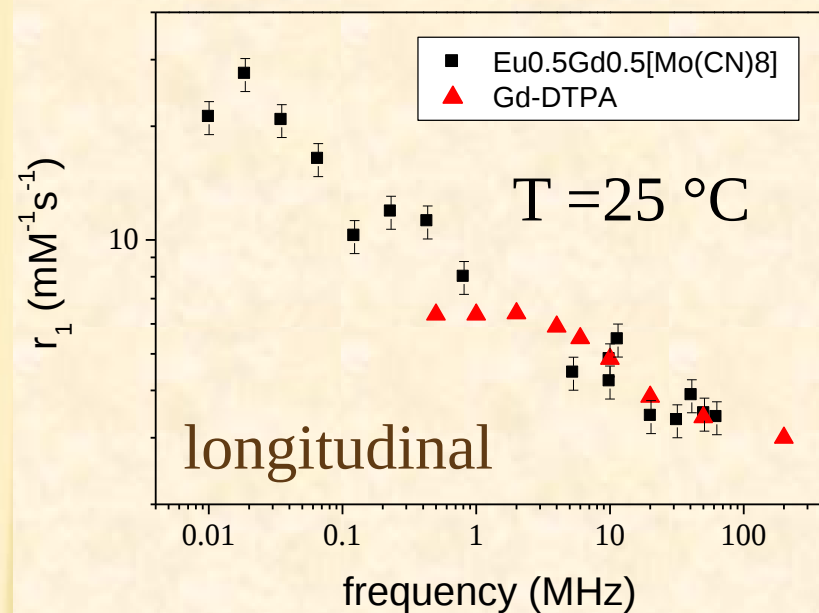


HRTEM image

Relaxometry of $\text{Gd}_{0.5}\text{Ln}_{0.5}[\text{Mo}(\text{CN})_8]/\text{chitosan}$ ($\text{Ln} = \text{Eu}, \text{Tb}$) Nanoparticles



$$r_{ip} = [(1/T_i)_{\text{meas}} - (1/T_i)_{\text{dia}}] / c \quad i=1,2$$



Luminescence of $\text{Gd}_{0.5}\text{Ln}_{0.5}[\text{Mo}(\text{CN})_8]/\text{chitosan}$ ($\text{Ln} = \text{Eu}, \text{Tb}$) Nanoparticles

$\text{Gd}_{0.5}\text{Tb}_{0.5}[\text{Mo}(\text{CN})_8]/\text{chitosan}$

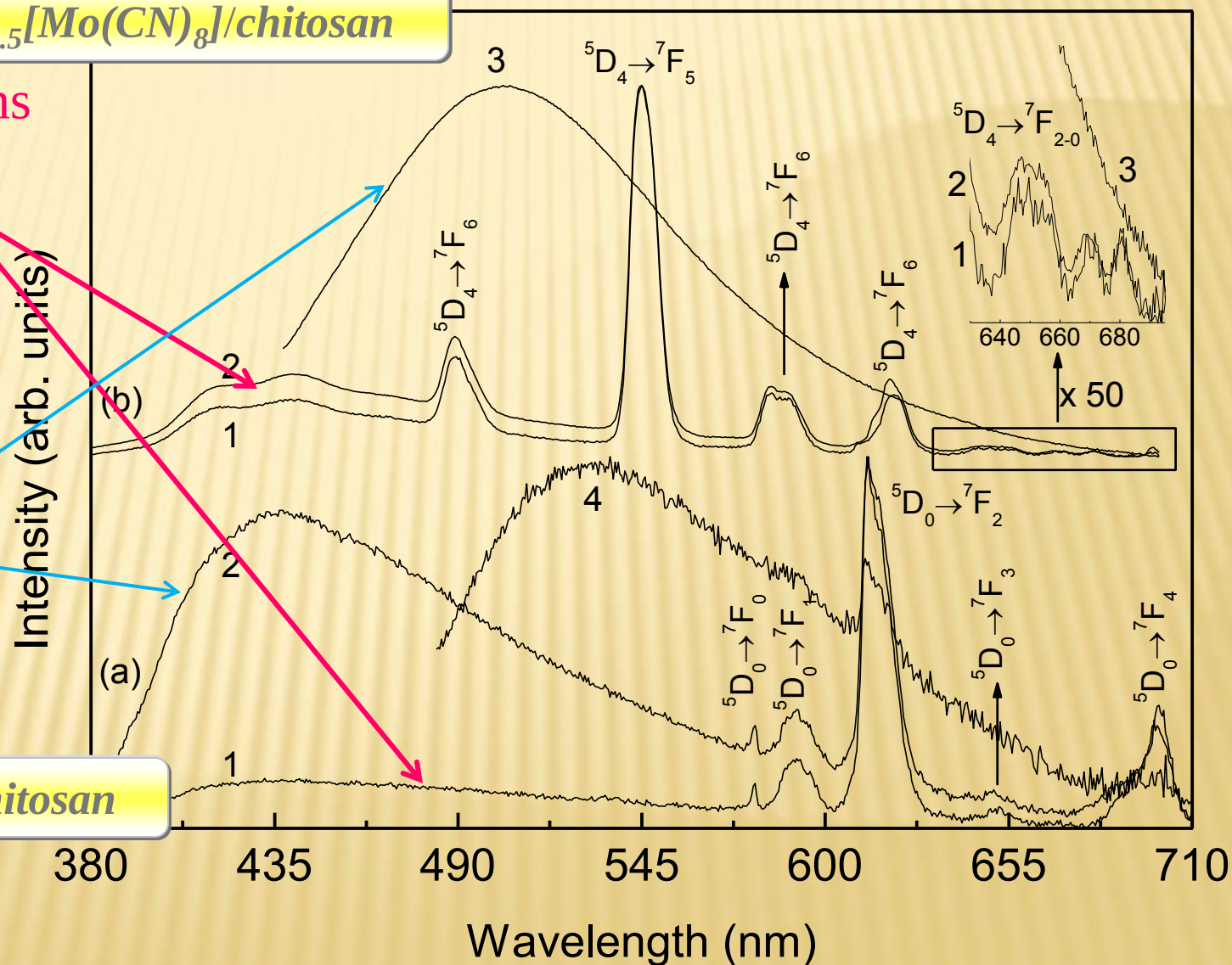
Excitation wavelengths
280-340 nm:

$^5\text{D}_0 \rightarrow ^7\text{F}_{0-4}$
(Eu^{3+})

$^5\text{D}_4 \rightarrow ^7\text{F}_{6-0}$
(Tb^{3+})

Excitation
wavelengths
350-464 nm:

$\text{Gd}_{0.5}\text{Eu}_{0.5}[\text{Mo}(\text{CN})_8]/\text{chitosan}$



Localization of $\text{Gd}_{0.5}\text{Eu}_{0.5}[\text{M}'(\text{CN})_m]_z/\text{chitosan}$ in living colon and pancreatic cancer cells by confocal microscopy.

Incubation:
6h at 37 °C
100 ng mL⁻¹

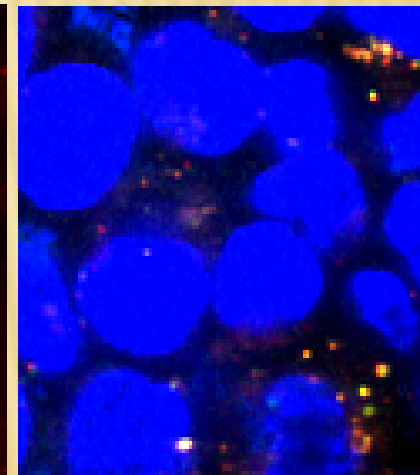
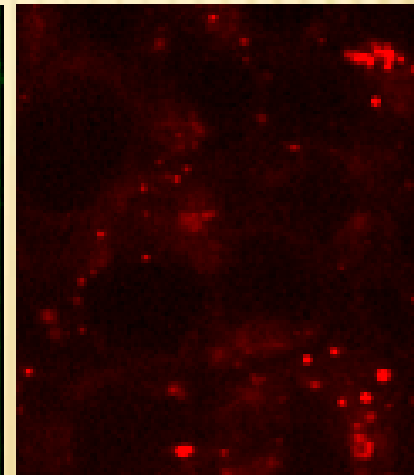
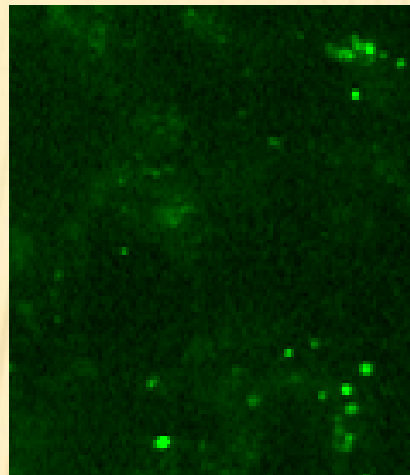
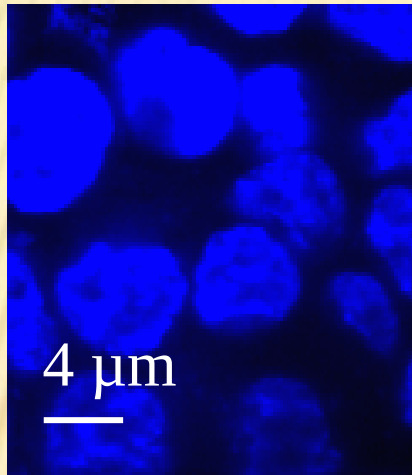
Nuclei

Sample 3
Excitation at 489 nm

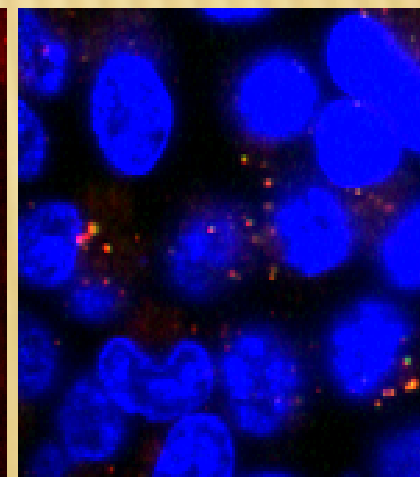
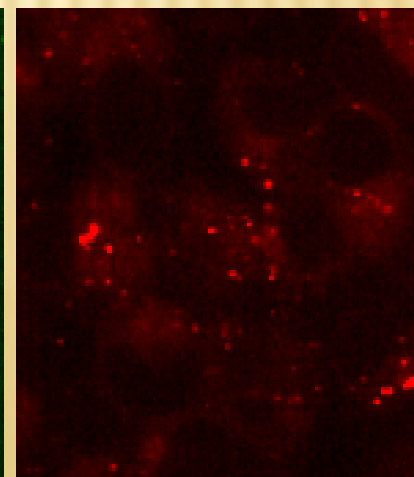
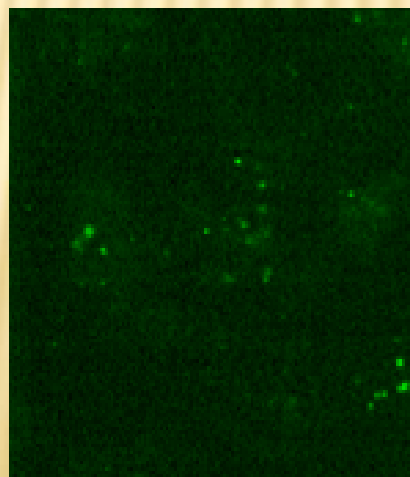
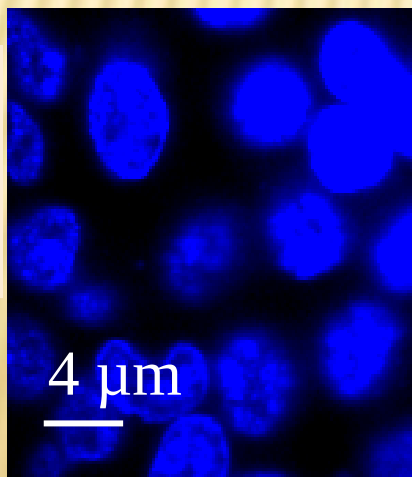
Lysosomes

Merged

HCT-116
colon



Capan-1
pancreatic
cancer cells



Toxicity Tests

4 days,

Concentration:

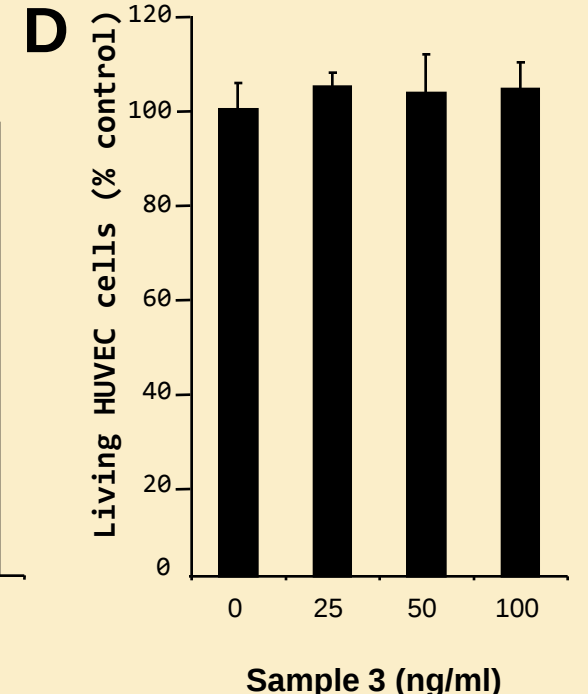
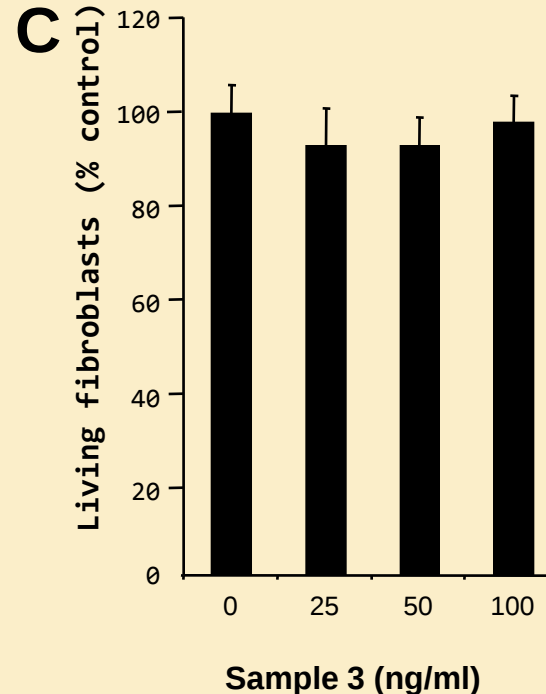
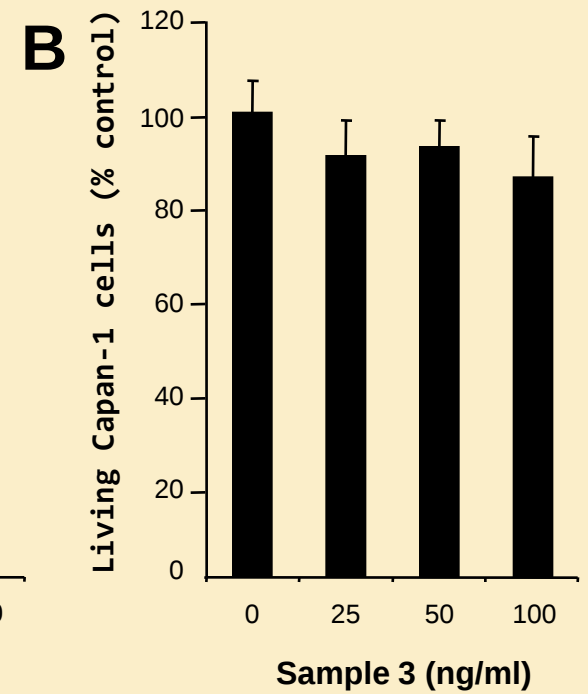
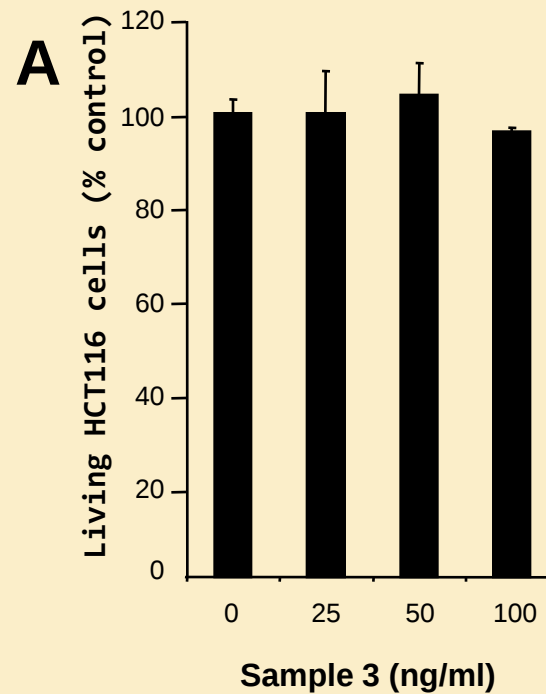
(25, 50 and 100 ng mL⁻¹)

(A) Human colorectal cancer cells (HCT116),

(B) human pancreatic cancer cells (Capan-1)

(C) human fibroblasts

(D) human umbilical endothelial HUVEC cells

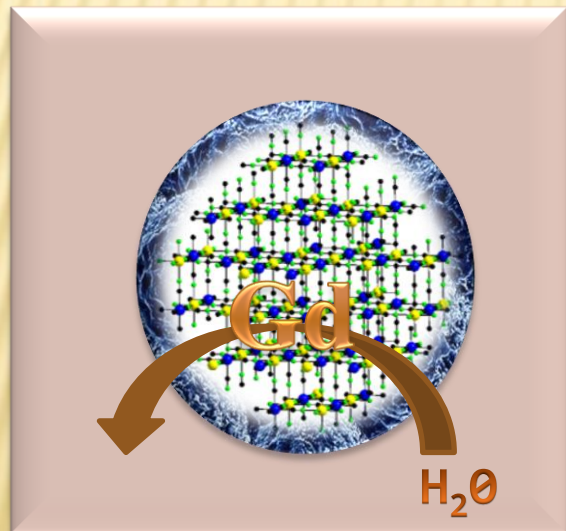


Summary:

Coordination Polymer Nanoparticles



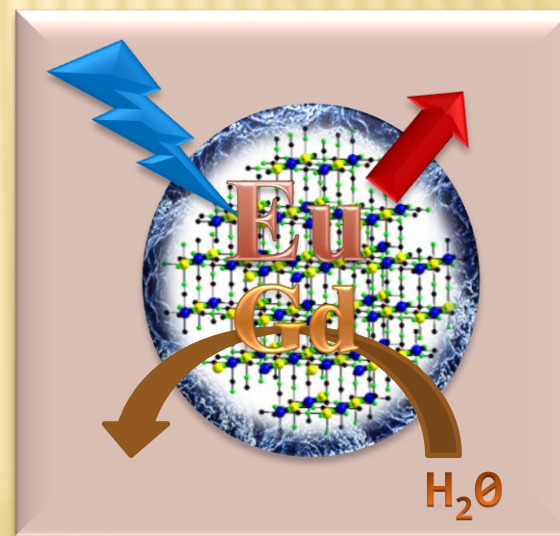
$\text{Gd}[\text{Fe}(\text{CN})_6]/\text{chitosan}$



Relaxivity



Multifunctional
 $\text{Gd}_{0.5}\text{Ln}_{0.5}[\text{Mo}(\text{CN})_8]/\text{chitosan}$
(Ln = Eu, Tb)



Relaxivity, Luminescence

Our group:

ICG-CMOS, Montpellier



Pr. Ch. Guérin

Dr. Y. Guari

E. Chelebaeva

Dr. B. Folch

Dr. A. Tokarev

C. Reibel

Our partners:

Dr. A. Caneschi, Dr. C. Sangregorio INSTM, Florence, Italy

Dr. C. Luna UANL, Mexico

Dr. A. Lascialfari, K. Tangawel, M. Marione, IFGCB, Milano, Italy

Pr. Luis Carlos, Dr. R. Fereira CICECO, Portugal,

Dr. N. Baril LNIA, Marseille, France

Dr. M. Garcia, Dr. M. Gary-Bobo IRC, Montpellier, France

Dr. K. Molvinger, MACS, ICG, Montpellier