

# FUNCTIONALIZATION OF ELECTRODE SURFACES WITH THREE-DIMENSIONAL NETWORKS OF ELECTROPOLYMERIZED GOLD NANOPARTICLES FOR BIOSENSOR DESIGN

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Research Team on Electroanalysis and Electrochemical (Bio)sensors



# METHODOLOGIES FOR NANOSTRUCTURING ELECTRODE SURFACES FOR BIOSENSING PURPOSES

i) TYPE OF ELECTRODE SURFACE

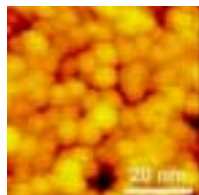
ii) NANOMATERIALS TO BE USED

iii) OTHER COMPOUNDS/MATERIALS

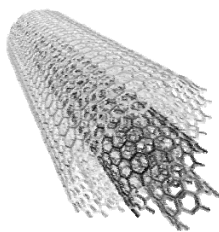
iv) PHYSICAL/CHEMICAL METHOD FOR SURFACE MODIFICATION

v) BIOMOLECULE TO BE IMMOBILIZED AND METHOD TO DO THAT

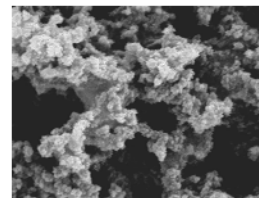
vi) ELECTROCHEMICAL REACTION TO TAKE PLACE



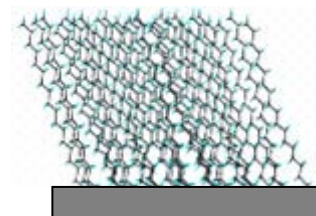
AuNPs



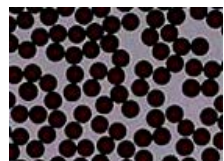
MWCNTs



CPs



SAMs



MNPs



Ab

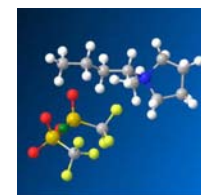
**Materials**



E

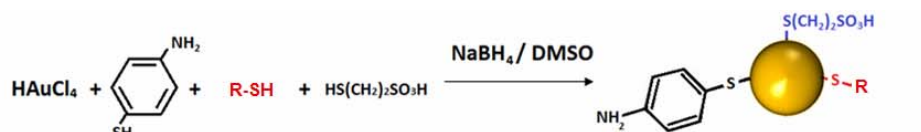
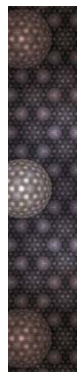


DNA

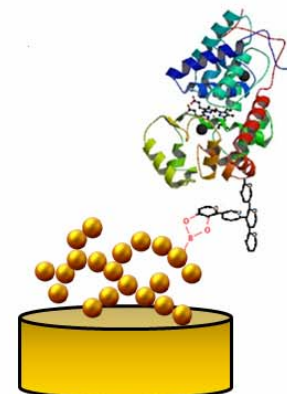
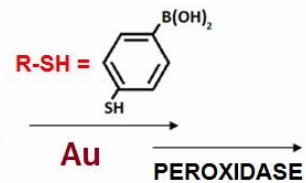


RTILs





Electropolymerization

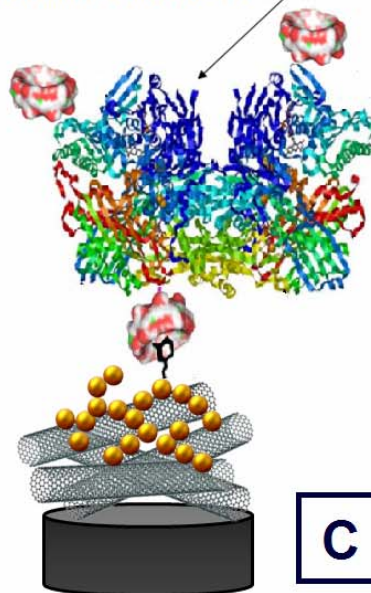


A

R-SH = 1-ADAMANTANETHIOL

GCE/SWNT

XANTHINE OXIDASE-CD

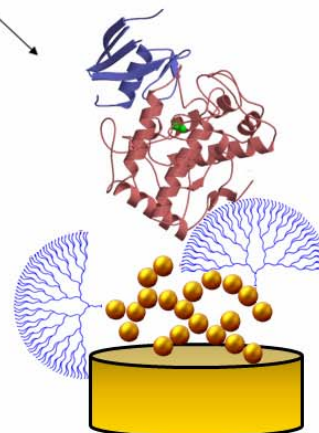


C

R-SH = PAMAM G4 CYSTEAMINE CORE DENDRON

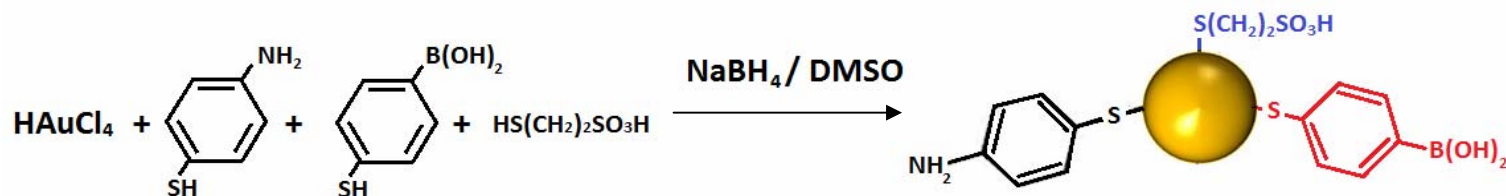
Au

TYROSINASE

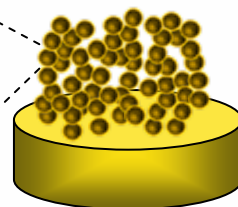


B

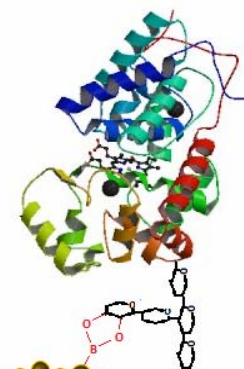
# GOLD NANOPARTICLES-BASED NANOSTRUCTURED POLYMERIC NETWORK



ELECTRODE  
Electropolymerization



HRP

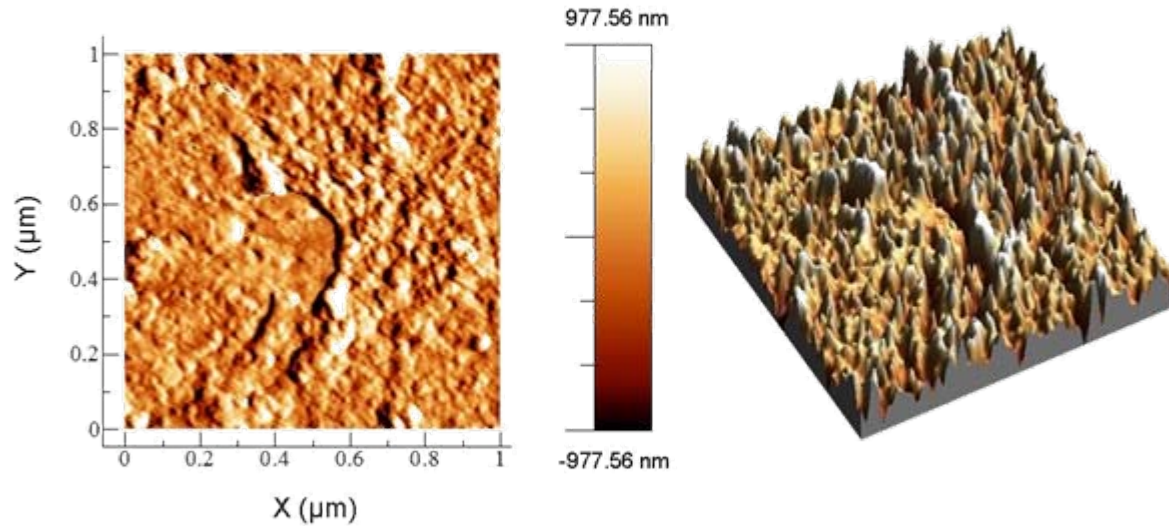


HRP wiring



mediatorless  
 $\text{H}_2\text{O}_2$  biosensor

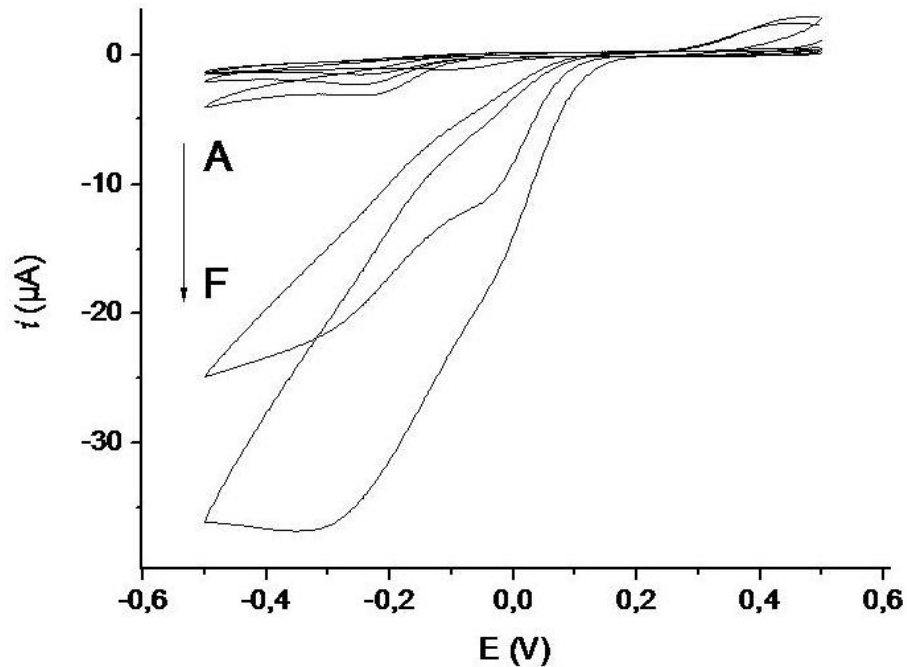




- average thickness: 984 nm
- nanoholes and AuNPs-based protuberant structures represented 47.9% and 52.1% of the total surface area, respectively.
- maximum height of AuNPs-based protuberant structures 1.96 μm

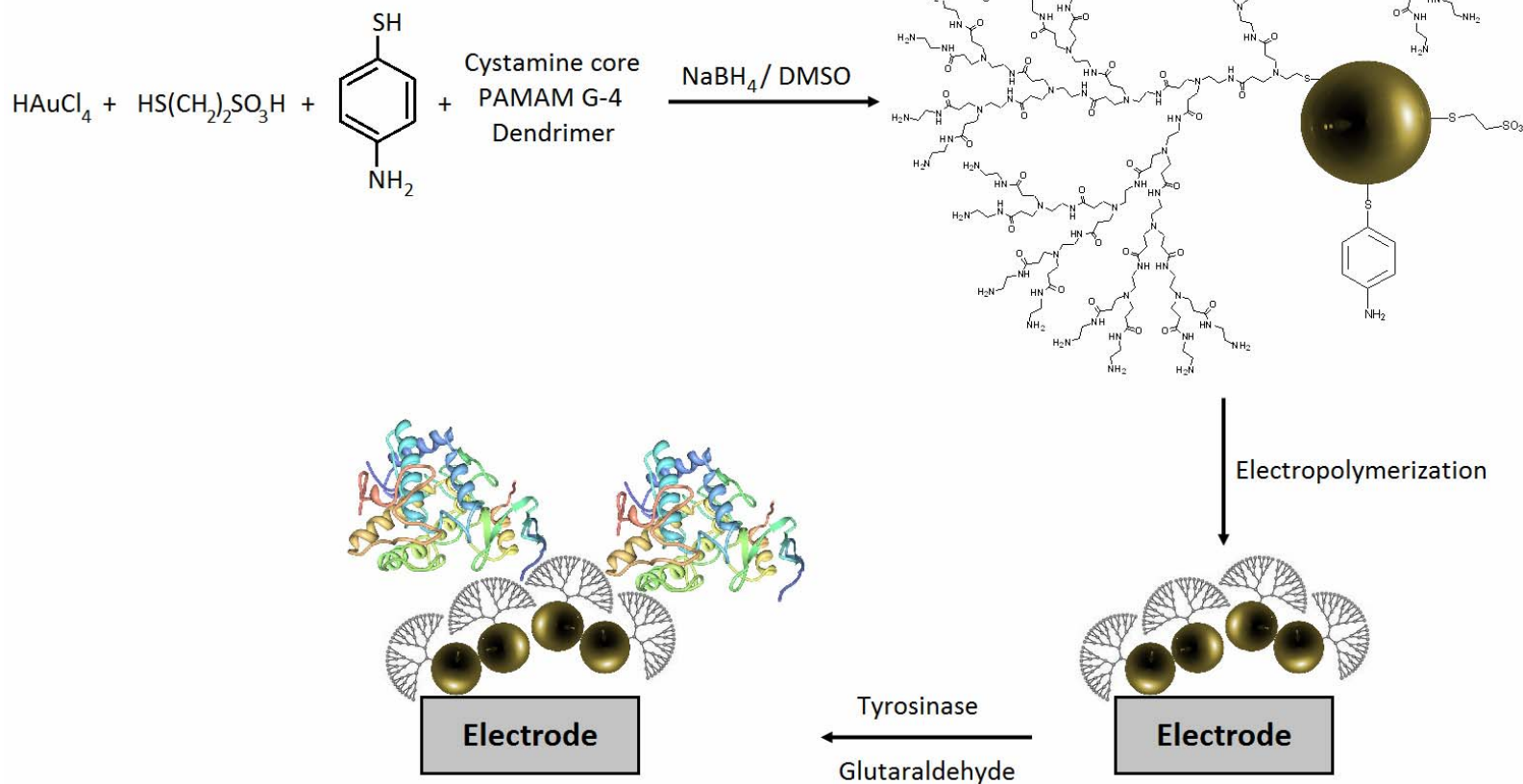


- A) HRP-Au
- B) HRP-polyAuNPs-Au
- C) HRP-Au + 1  $\mu\text{M}$   $\text{H}_2\text{O}_2$
- D) HRP-Au + 2  $\mu\text{M}$   $\text{H}_2\text{O}_2$
- E) HRP-polyAuNPs-Au + 1  $\mu\text{M}$   $\text{H}_2\text{O}_2$
- F) HRP-polyAuNPs-Au + 2  $\mu\text{M}$   $\text{H}_2\text{O}_2$

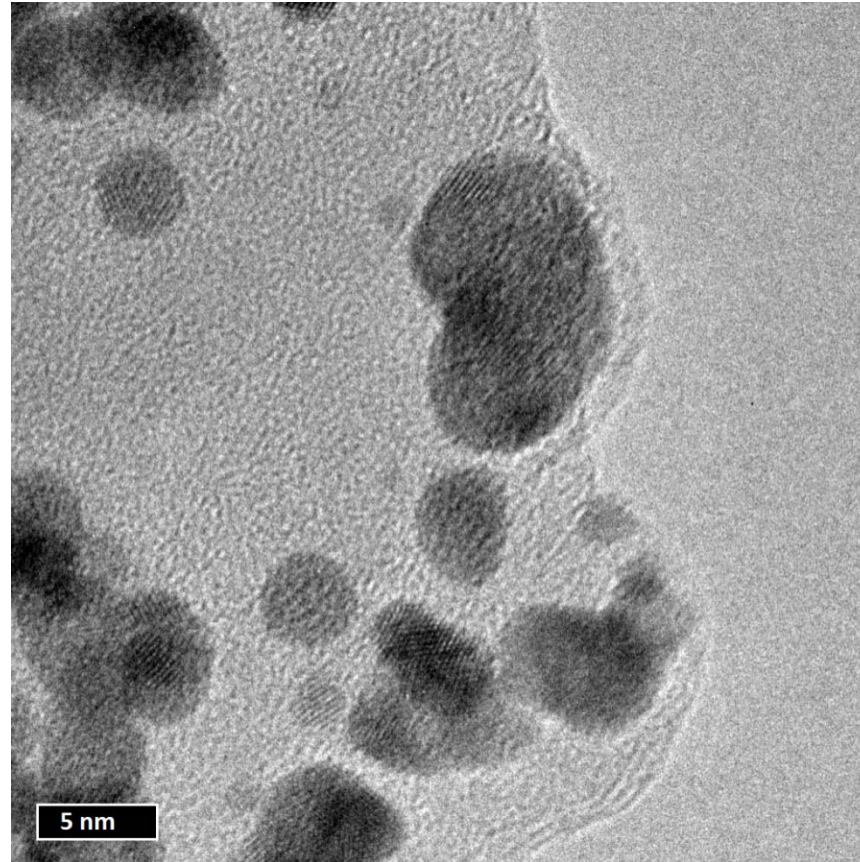


- Conclusions
- Improved electrocatalytic behaviour of HRP upon immobilization on the polyAuNPs-based conductive matrix
- Efficient electron transfer between the redox active site of the enzyme and the electrode surface



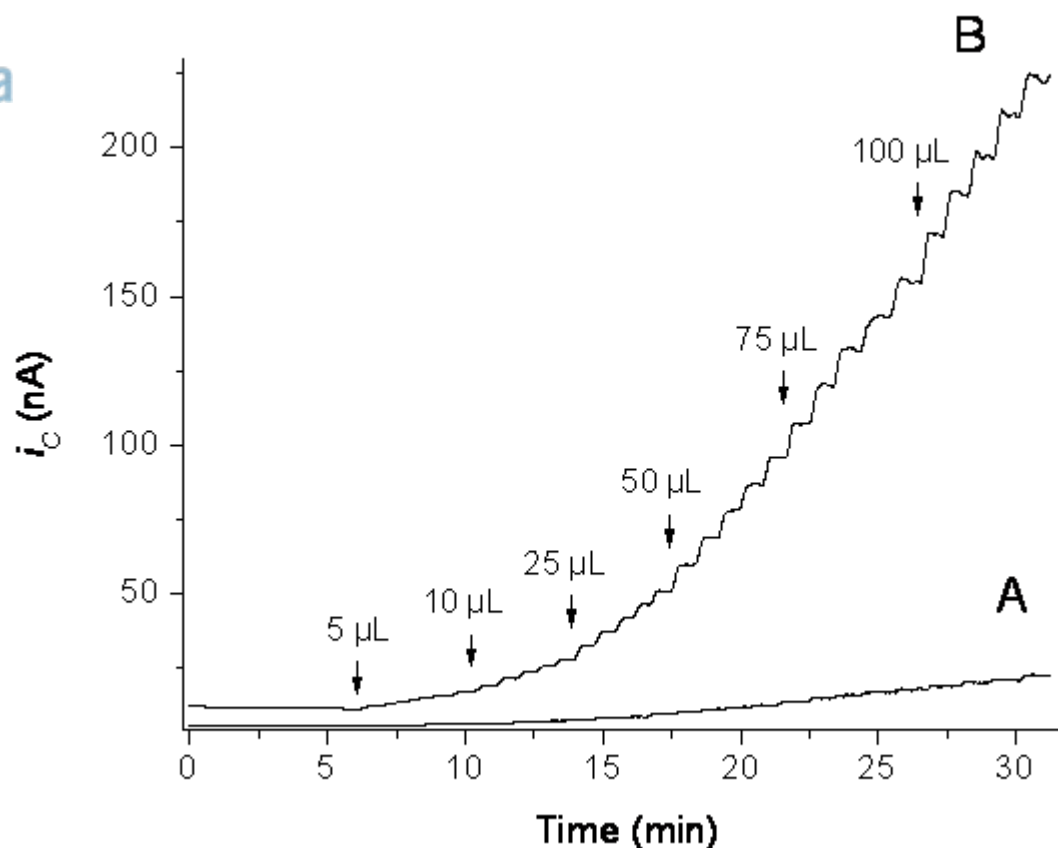






- Average diameter  $5.7 \pm 0.9$  nm
- PAMAM G-4 dendrimers: about 4.5 nm
- Larger than that reported for metal nanoparticles encapsulated into dendritic molecules





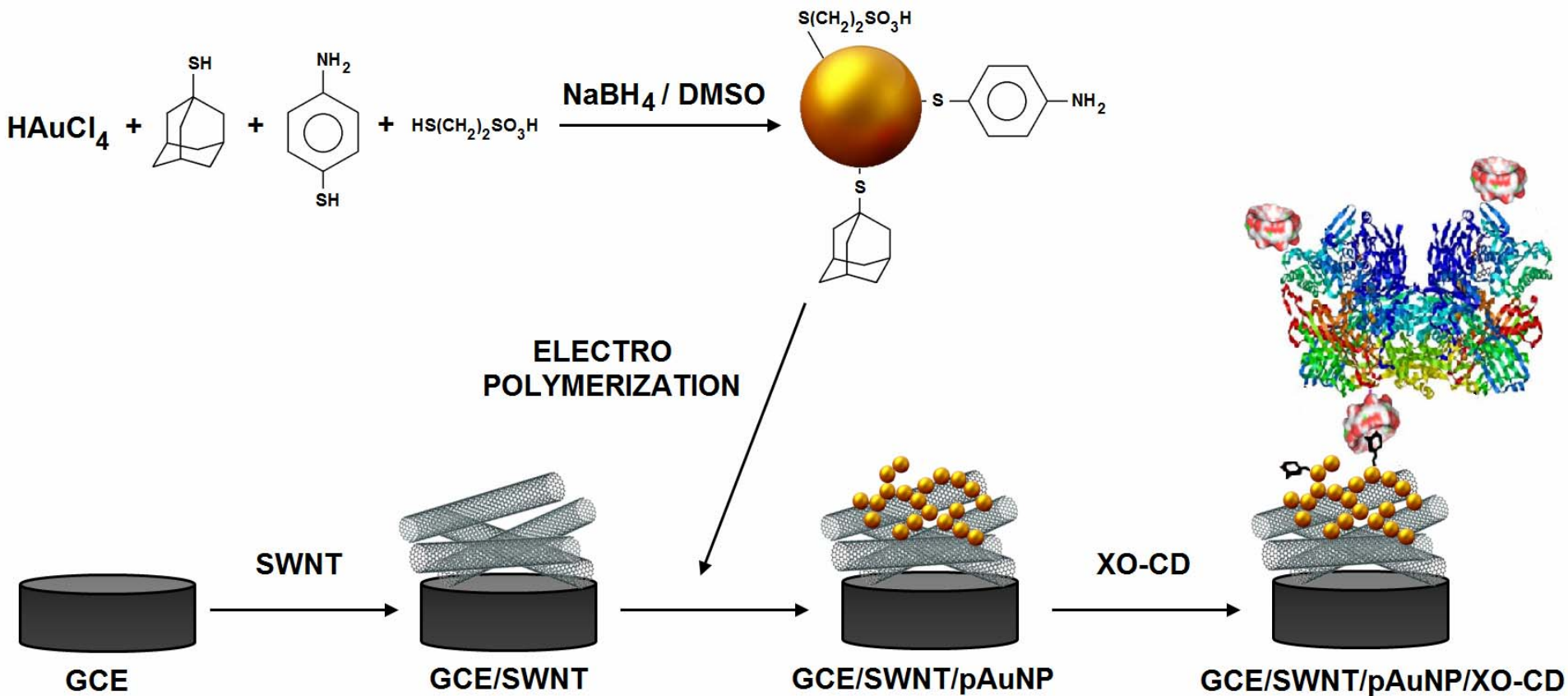
Tyr-functionalized *p*-aminothiophenol-coated non-nanostructured gold electrode (A) and electropolymerized PAMAM G-4 dendron-coated AuNPs-modified electrode (B)

### Analytical characteristics

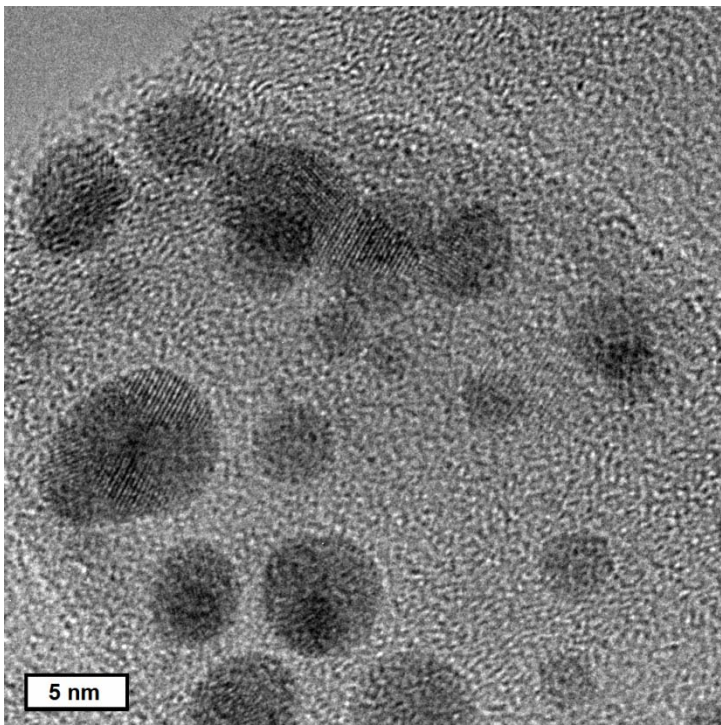
Detection limit: 20 nM; Range of linearity: 50 nM - 10  $\mu$ M.  $K_M$  = 21.9  $\mu$ M

RSD = 3.1% for 10 measurements of 100 nM catechol using the same electrode; electrode-to-electrode reproducibility RSD = 8.8% (n = 10)



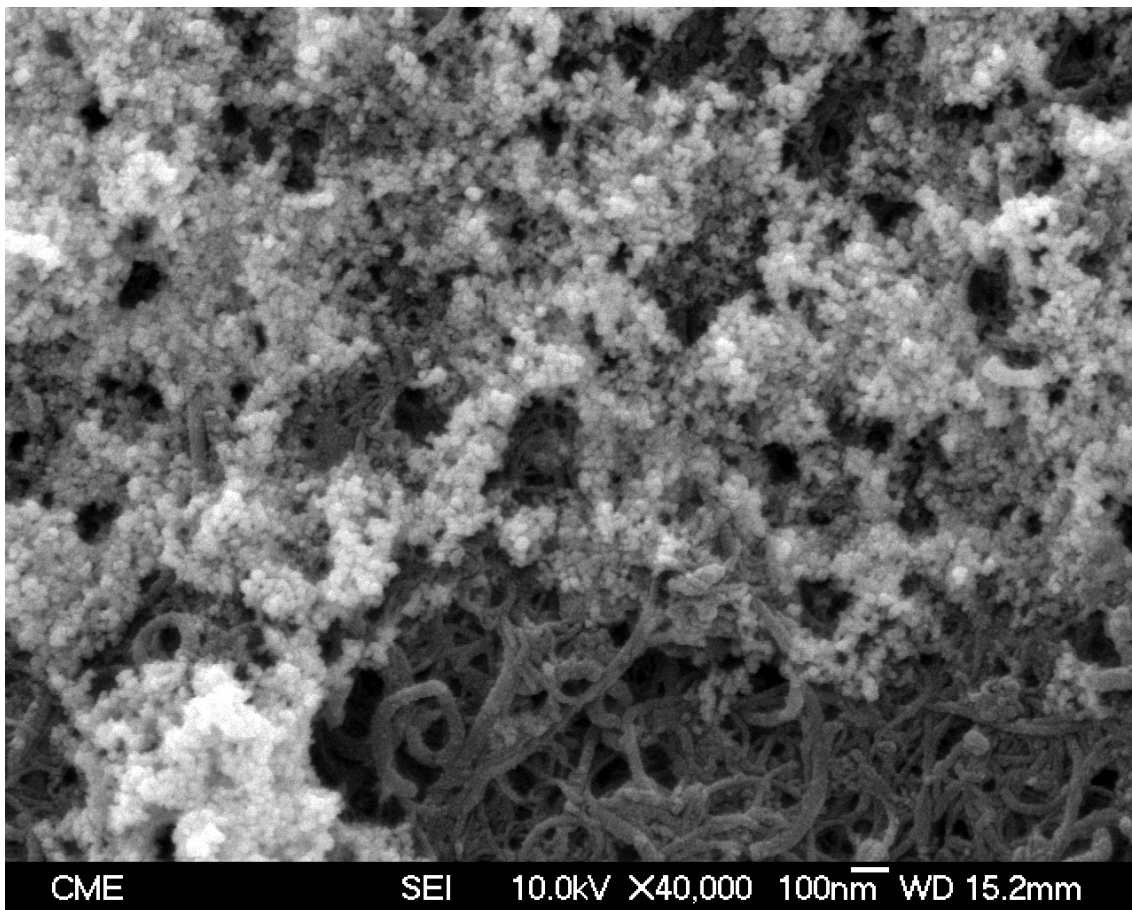




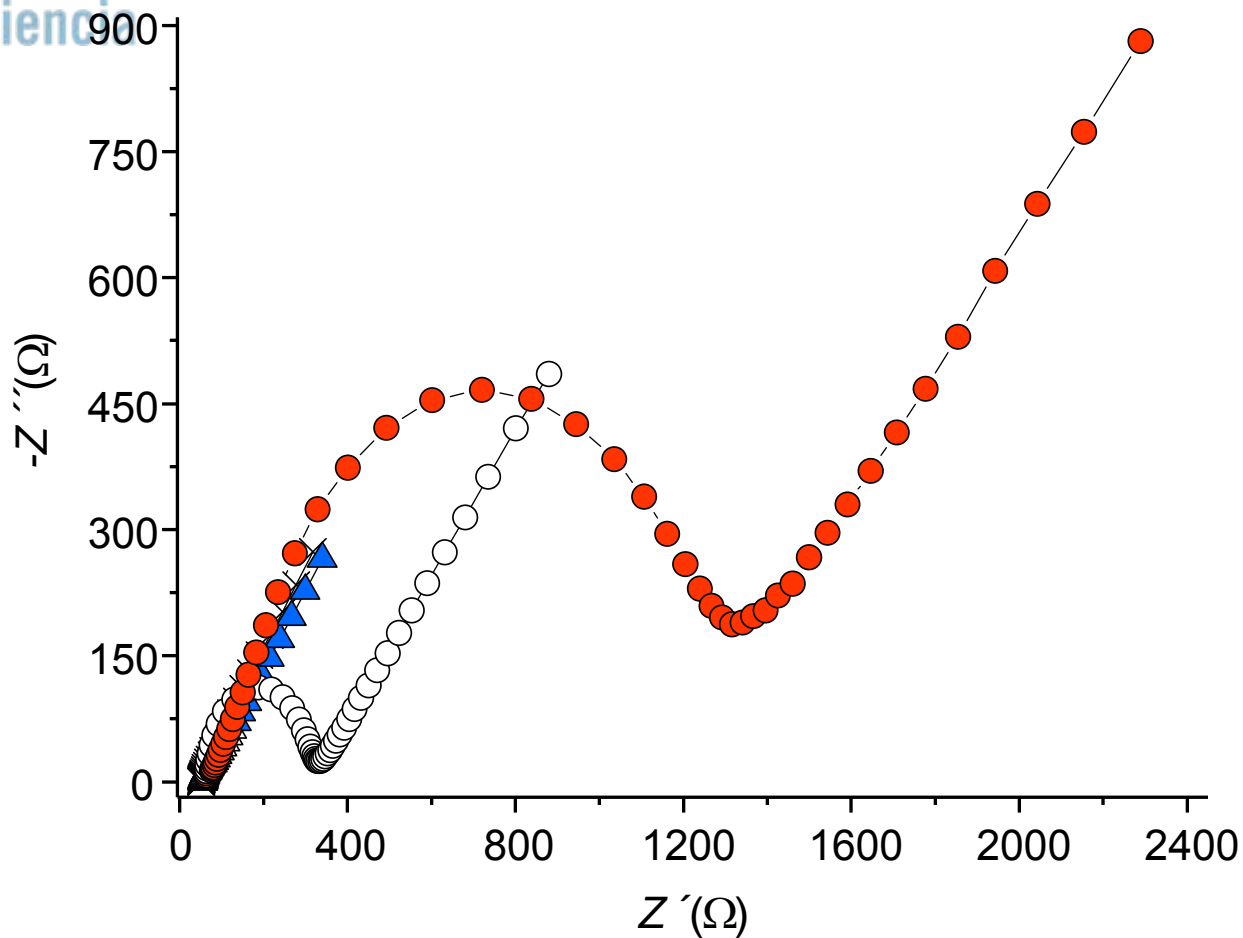


HRTEM  
 $4.9 \pm 0.8$  nm

SEM



CME SEI 10.0kV X40,000 100nm WD 15.2mm

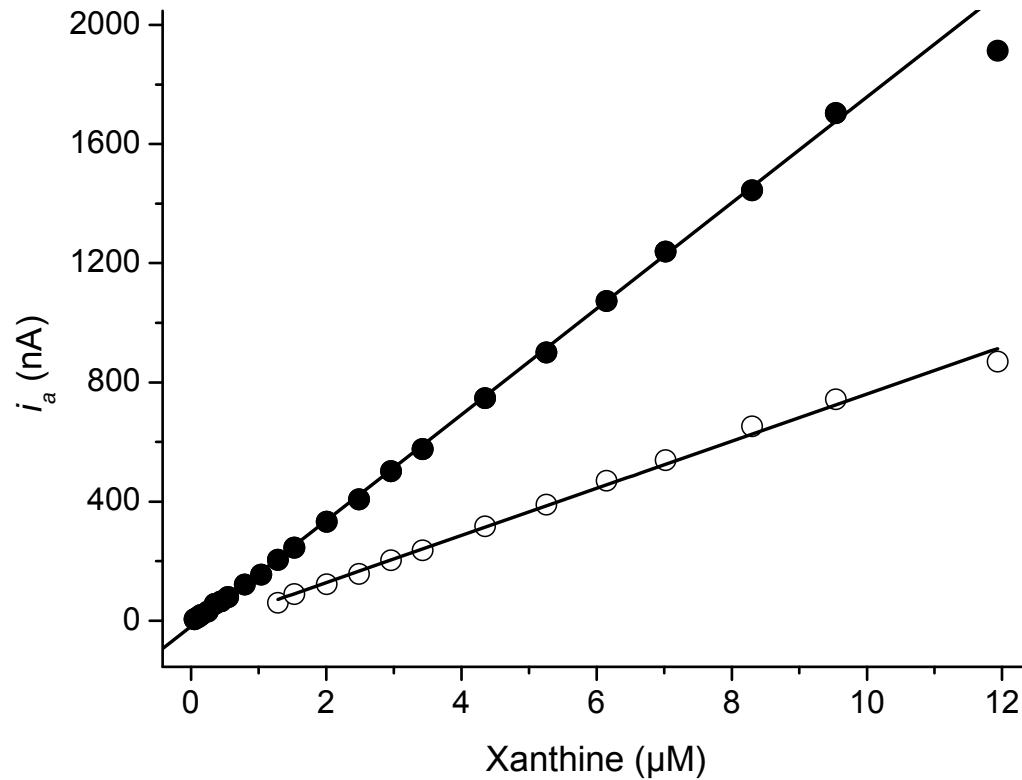


EIS, (○) GCE; (▲) GCE/SWNT; (X) GCE/SWNT/pAuNP; (●) GCE/SWNT/pAuNP/XO-CD  
5 mM  $\text{Fe}(\text{CN})_6^{3-/4-}$





(●) XO-CD; (○) native XO



Sensitivity: 4.71 A/M  
 $\text{cm}^2$  ( $A = 4.1 \text{ mm}^2$ )

Linear range: 50 nM – 9.5  $\mu\text{M}$   
Limit of detection: 40 nM



## CONCLUSIONS

The use of bisaniline-cross-linked nanostructured networks of metal nanoparticles constitutes an excellent strategy to design scaffolds for the successful immobilization of redox enzymes to prepare mediatorless amperometric biosensors with improved analytical performance.

### Acknowledgements

Financial support from the Spanish Ministerio de Ciencia e Innovación CTQ2011-24355 is gratefully acknowledged



**MUCHAS GRACIAS**